Timing of triazole-based spray schedules for controlling mungbean powdery mildew in Australia: a meta-analysis

Paul Melloy¹, Emerson Medeiros Del Ponte², Adam H. Sparks^{1,3}

¹ University of Southern Queensland, Center for Crop Health, Toowoomba, Queensland, 4350 Australia

² Departamento de Fitopatologia, Universidade Federal de Viçosa, Viçosa MG 36570-000 Brazil

³ Department of Primary Industries and Regional Development, 3 Baron-Hay Court, South Perth, 6151, Western Australia, Australia

Abstract

Powdery mildew (PM), caused by two fungal species, *Podosphaera xanthii* and *Erysiphe vignae*, is a yield limiting foliar disease commonly found in mungbean (*Vigna radiata*) cropping areas of eastern region of Australia. Effective control of PM afflicting mungbeans relies largely on fungicide applications, mainly of the triazole group. Uncertainty in the current fungicide spray schedule recommendations, which advise commencing with a spray at the first sign of PM, prompted this study to evaluate PM severity and crop yield data obtained from fungicide trials which tested spray schedules starting before (*Early*) or after (*Late*) first signs, applied singly or combined with a follow-up spray. A meta-analytic approach was employed to obtain estimates of the yield and PM severity difference between plots sprayed with specific triazole-based sprays schedules and untreated plots. From 26 trials, 15 and 14 met the criteria for inclusion in

the yield and PM severity analysis respectively. The schedule with the first spray starting at first sign with a follow-up spray 14 days later, resulted in significantly lower disease severity compared to all other schedules, however, the yield protected was only numerically higher and not statistically different compared to: single-spray at first sign, single-spray *Late* or two-spray starting *Late*. While PM severity was lower in the *Early* sprayed plots compared to untreated plots, yield did not differ. These findings support the current recommendations and provide evidence that the first spray could be delayed up to a week and save an additional spray that may be unnecessary, thus reducing direct fungicide costs as well as indirect costs due to fungicide resistance.

Keywords: *Podosphaera xanthii*, *Erysiphe polygoni*, *Erysiphe vignae*, *Vigna radiata*, demethylase inhibitors, network meta-analysis

Introduction

Mungbean [*Vigna radiata* (L.) Wilczek] is a pulse crop primarily grown in Southeast Asia for human consumption either as raw pods, grain or added to meals after sprouting as an affordable source of protein (Lambrides and Godwin 2007). The dried grain can also be ground into a protein-enriched flour for uses in noodles, biscuits and cakes (Chankaew et al. 2013). Even though mungbean was first brought to Australia in the 1930s for use as a forage or green manure crop, it is still considered a relatively new commercial crop. In

2011, a cost-benefit analysis valued the return on investment for mungbean research at 18:1 (Australian Mungbean Association n.d.). Since then, high value export markets and new varieties with improved yields have increased profitability and the crop has been more widely adopted. Between 35 and 86.4 thousand hectares of dried beans are now planted annually in Australia (Clarry 2016). Today, mungbean is predominantly grown in southern Queensland and northern New South Wales as a short season summer legume crop. In 2019, approximately 90% of Australian mungbean was grown for export which received attractive returns of up to \$1300 AU per tonne (Queensland Government 2019).

Mungbean potentially yields up to 3 tonnes per hectare (Thomas et al. 2004) but the average Australian farm yields less than 1 t ha⁻¹ (Chauhan and Williams 2018) with significant temporal and spatial variability, suggesting the urgent need for research efforts targeting production constraints. Abiotic constraints, insects and competition from weeds contribute to mungbean yield losses, in addition to the occurrence of diseases caused by fungi, bacteria and phytoplasmas (Wood and Easdown 1990; Conde and Diatloff 1991; Thakur and Agrawal 1995; Fuhlbohmer et al. 1996, 2013; Wilson et al. 2001; Noble et al. 2019)

Among the fungal mungbean diseases in Australia, powdery mildew (PM), caused by obligate biotrophic fungi *Podosphaera xanthii* (syn. *Podosphaera fusca* and *Erysiphe polygoni*) and *Erysiphe vignae* (Kelly et al. 2021), can reduce yield by 40% in susceptible

cultivars grown under disease-conducive conditions (Lambrides and Godwin 2007). To date, ascospores produced in chasmothecia (sexual stage) have not been reported in Australia; hence the asexual airborne conidial spores that are produced abundantly on live hosts and dispersed by wind currents, are considered the primary inoculum for epidemics while the alternate host or hosts remain unknown. Conditions for infection by PM pathogens are optimal when and temperatures range between 22 °C and 26 °C (Pérez-García et al. 2009; Kelly et al. 2017). Powdery mildew is a polycyclic disease that develops quickly; the time from conidial germination to newly producing conidiophores disseminating conidia (a monocycle) can be as short as five days (Sparks and Kelly 2017). The characteristic white powdery fungal growth on the infected leaves negatively affects photosynthesis, leading to reduced yield quantity and quality (Pérez-García et al. 2009). In severe cases leaves become chlorotic, which can lead to premature leaf senescence.

Powdery mildew management involves the integration of cultural, genetic and chemical practices (Pandey et al. 2018; Sparks and Kelly 2017). In southern Queensland and northern New South Wales, sowing of mungbean crops is recommended between late spring (November) and mid-summer (January) to avoid the cooler autumn temperatures that favor the disease (Australian Mungbean Association n.d.). While loci conferring quantitative resistance to PM in mungbean have been incorporated in mungbean breeding lines (Humphry et al. 2003; Zhang et al. 2008; Pandey et al. 2018; Douglas 2008); ongoing breeding efforts in Australia have focused on resistance to

bacterial diseases given the feasibility to use fungicides for PM control. The recently released Opal-AU cultivar is rated as moderately susceptible (MS) to moderately resistant (MR) and is considered to contain the best resistance to PM (Douglas and McIntosh 2020) followed by Jade-AU (MS) and Green Diamond (Australian Mungbean Association 2016). Losses up to 32.7% have been recorded in Jade-AU (Thompson 2016). However, cultivar resistance alone is not sufficient to control PM, hence fungicide applications are necessary when conditions for the disease are conducive.

Field trials conducted in eastern Australia over the past ten years have evaluated fungicide efficacy with a range of active ingredients and spray timings. The trial results are available only as grey literature reports (Kelly et al. 2017; Thompson 2016; Northern Growers Alliance 2013d, 2013c, 2013a, 2013b). In general, the best control of PM for yield protection, has been achieved with a first application at the first sign of the disease, with a follow-up application two weeks later if conditions are favourable for disease development (Sparks and Kelly 2017; Thompson 2016). However, these results have been inconsistent, possibly due to environmental or trial-specific conditions as well as lack of statistical power in individual trials. Additionally, a fair comparison among several treatment options was not possible due to different combinations of treatment settings between the trials, such as fungicide active ingredient, number of sprays and spray timing. These issues can be dealt with by combining results from the primary studies to obtain estimates of both the size and uncertainty of the treatment effects via a systematic

review with meta-analysis (MA). The method has become standard in plant pathology for summarizing data from uniform fungicide trials. Typically, MA models are fitted separately to data for each treatment against a control group (Madden and Paul 2011). Alternatively, a single model can be fitted in the form of a network of treatments where both direct and indirect evidence is used in the calculation of treatment effect sizes. The latter approach is called a network meta-analysis, one of the several meta-analytic methods that has found use for multiple treatment comparisons in plant disease management due to its estimates being considered more precise in some contexts (Madden et al. 2016).

The recently launched mobile app '*PowderyMildewMBM*' (Diggle 2019) is a decision support tool that provides a cost-benefit analysis to assist growers evaluate whether applying fungicide to treat PM would be profitable given their weather conditions and the estimated value of the mungbean crop. More robust and reliable estimates of yield responses to different fungicide spray schedules obtained using meta-analytic modelling could be important for validating the assumptions the app makes about the timing of fungicide applications, such as avoiding prophylactic sprays.

Two questions were addressed in our study: i) when in time, relative to PM first signs (prior, during or after), should the first fungicide spray be applied on mungbean to maximize yield protection, ii) given those timings at which the first spray was made is a second spray worthwhile yield protection? To answer these questions, we systematically

reviewed all sources of fungicide trial data for PM control in the eastern Australian mungbean production scenarios that we were able to obtain and conducted a meta-analysis of the effect of different sprays schedules on mungbean yields.

Materials and Methods

Data source and inclusion criteria

As of August 2020, no peer-reviewed study on the efficacy of fungicide on powdery mildew in mungbean was found to be conducted in Australia after searching article databases, *Web of Science*, *Scopus* and *Google Scholar* with the search string: “mungbean AND powdery mildew AND fungicide” and filtering the results by country of publication. On the other hand, we were able to find and gather data of the fungicide effect on PM intensity and mungbean yields from University of Southern Queensland owned research databases and collaborating institutions including Northern Growers Alliance and the Queensland Department of Agriculture and Fisheries. The data were obtained in a variety of formats and accompanying information. While raw data were available for thirteen trials, others reported only the means of each treatment.

The collated field trials not meeting the following criteria were excluded: i) Field trial conducted in Australia ii) record of the date when powdery mildew was first observed;

iii) measure of disease severity at the end of the growing season; iv) fungicide application dates; v) the fungicide active ingredient(s); vi) fungicide dose; vii) and crop yield.

Treatments with fungicide active ingredients evaluated in fewer than eight trials were discarded from the analysis. This resulted in the retention of two DMI fungicides (tebuconazole and propiconazole) in the dataset and the initial set of 26 trials was reduced to 17 trials. These 17 trials were further reduced in number as some trials did not report either yield or PM severity variance along with the mean, which was required for the meta-analysis, leading to two datasets for each meta-analysis. Two trials, 1112/01 and 1516/03, did not report grain yield variance and three trials, 1516/01, 1516/02, 1516/03, did not report severity variance. Therefore, data from 15 trials were used for the mungbean yield analysis and 14 trials for PM severity analysis (Table 1). All trials included in this analysis were conducted in a randomised complete block design. Further trial details can be found in table 1 of the supplementary materials.

Treatments of interest

In this study, we were interested in testing whether variations in spray schedules based on the use of a triazole fungicide (tebuconazole and/or propiconazole) are more beneficial than the recommended practice. The '*Recommended*' practice referred to a single fungicide spray schedule starting one to three days following the first sign of disease. After inspecting the trial data and meta-data that met our inclusion criteria, four other

spray schedules were defined based on a combination of spray timing and one or two follow-up sprays (coded with the suffix +). A total of five triazole-based spray schedules were then defined: *Recommended* ($n_{yield} = 27$, $n_{severity} = 28$); *Recommended+* ($n_{yield} = 40$, $n_{severity} = 41$); *Early*, single spray prior to disease detection ($n_{yield} = 13$, $n_{severity} = 13$); *Late*, single spray from seven and 13 days after the first sign of disease ($n_{yield} = 17$, $n_{severity} = 15$); and *Late+* ($n_{yield} = 19$, $n_{severity} = 13$). There were no spray schedule treatments that began between four and six days after the first sign of powdery mildew. Given the small number of trials, the *Early+* ($n_{yield} = 5$, $n_{severity} = 5$) was excluded from the meta-analysis.

Response variable and meta-analytic procedures

Following previous meta-analyses to evaluate the effect of fungicides on crop yield (Paul et al. 2008), we were interested in obtaining estimates of the yield and disease severity difference between spray schedules and the untreated control. A preliminary inspection of mean grain yields at the trial level suggested that a normal distribution could be assumed, as was also the case for PM severity. Grain yield means of each fungicide treatment, expressed as metric tons per hectare, were either obtained directly from the summaries of trial reports, or summarised from the raw data when available. PM severity was assessed on an ordinal scale from one (no signs of the pathogen) to nine (plant totally infected and dropping leaves). The sampling variance was also calculated

from the raw data when available or, when trial data was obtained from trial summary results, the least squares statistic using the method described in Ngugi et. al. (2011).

A network meta-analysis, also referred to “two-way unconditional linear mixed model” was chosen because it assumes correlations between treatments within the same trials and makes weighted comparisons between trials even if all treatments are not present within each included trial (Paul 2008; Machado 2017). The model is given by

$$Y_i \sim \text{Norm}(\mu + \delta, \Sigma + S_i^2),$$

Where Y_i , the response, is either a vector of the grain yield measurements (tonnes per hectare) or a vector of PM severity ratings, for the i th trial over the total number (K) of trials ($i = 1, \dots, K$). Y_i is assumed to be estimated from a normal distribution with μ the estimated weighted means of treatments across all trials; δ is the estimate for each moderator level, in this case ‘*spray schedule*,’ corresponding to each fungicide treatment level; Σ , a 6 x 6 unstructured matrix of treatment variances between trial; and S_i^2 a variance-covariance matrix. Weighted estimates of this random effects model were calculated using the inverse of the model-implied variances (Viechtbauer 2010). The model was fit so ‘*spray schedule*’ treatments were added as correlated random effects within each trial where the variance co-variance matrix of the random effects was specified as ‘unstructured’. This allows the random effects for each spray schedule treatment to have different variances, which are also allowed to be correlated. Each mean

and variance entry were given an individual '*id*' variable, which was incorporated as a random effect to the model. Percentage of powdery mildew control ($\bar{C}$) was calculated by

$$\bar{C} = \frac{\mu_c + \mu_s}{\mu_c} \times 100$$

Where μ_c is the mean disease severity estimate in the no-spray control, μ_s is the severity estimate attributed to the spray schedule.

The model was fitted in the R statistical software environment, version 4.0.1, (R Core Team 2020) using the *maximum likelihood* parameter and '*optim*' optimizer with the '*metafor*', version 2-4.0, R package (Viechtbauer 2010). Linear contrasts of the spray schedule moderator estimates and variances for grain yield were used to obtain the mean difference and associated 95% confidence intervals between each spray schedule. These mean differences were used to infer the optimum fungicide spray strategy for maximizing mungbean grain yields.

Results

Trial characteristics and overall summary. Characteristics of the trials which met the study inclusion criteria were summarized in Table 1. Planting dates for the trials ranged from late December to the middle of February. The first signs of powdery mildew were observed as early as February 28 and as late as April 12. As expected, PM severity was higher, and the mean grain yields lower in the unsprayed control treatments compared

to when sprays were applied. The mean yields at the trial level ranged from 289.4 kg ha⁻¹ to 2045 kg ha⁻¹ in the control, while in the spray schedules they ranged from 275.9 t ha⁻¹ to 2526.3 kg ha⁻¹ (Table 1).

In general trials that yielded below 700 kg ha⁻¹ showed no yield benefit from any fungicide application and two-thirds of the trials produced yields which were on average higher in the fungicide treated plots compared with the no spray control (Table 1).

Meta-analytic estimates: The end of season PM severity was significantly lower in all fungicide spray schedules compared to no spray controls. The analysis estimated the *Recommended+* spray schedule provided the highest efficacy for controlling PM (Table 2), significantly lowering PM severity compared to all other spray schedules. The other spray treatments still provided effective control of PM ($P < 0.0001$) and did not differ significantly from each other with exception to the *Late+* spray schedule which was significantly higher than a single *Late* application ($P = 0.003$).

Despite the fungicide effect on PM severity, the spray schedule effect on mungbean yields were somewhat similar. All spray schedules but the *Early* ($P = 0.946$) resulted in a significant yield increase relative to the unsprayed controls (Table 3). For those spray schedules, the difference in yield relative to control ranged from 130.1 to 189.1 kg ha⁻¹, with the highest estimated yield increase produced by *Recommended+* followed by *Recommended*, *Late* and *Late+* (Table 3). A Wald-test of moderators indicated a disease pressure in the no spray control was significant ($P = 0.047$) when improving yield

protection estimates for each spray schedule. However due to low replication between some disease pressure categories and no *Late+* spray schedules observed in low disease pressure trials, further analysis was not pursued.

An inspection of log-likelihood profile plots showed the model for each random interactive term provided a reasonable estimate of τ^2 , with no indication the variables were over-fit.

Discussion

The current advice provided to eastern Australian mungbean growers for when to commence fungicide applications for controlling powdery mildew is to spray at the first sign of the disease (Thompson 2016; Sparks and Kelly 2017). Our meta-analysis corroborates the current advice but also suggests that if the first spray is delayed by one to two weeks mungbean yields would still be protected given the statistically similar results of the *Late* categorized spray schedules to the *Recommended* practice despite appearances the fungicide may have influenced disease severity. This is good news for mungbean growers who lack time and resources to scout crops regularly for disease or for situations when logistic issues prevent growers from getting machinery in the paddock immediately after PM is spotted. This flexibility to start a spray program permits growers to consider better planning for their integrated pest management by timing sprays in combination with other chemicals and thus reduce costs with spray application. However, if the grower decides to apply a second spray, consideration should be given

to the fungicide withholding periods, which in Australia require the second application to be applied at no earlier than 10 - 14 days after the first application, and not within 21 days of harvest (APVMA 2014). Time permitting, a second spray after the first “Late” application may not be necessary, as suggested by our results. More research is needed to evaluate whether crop age at the first sign of disease may have on crop yield reductions given spray schedule interventions.

According to the Australian Mungbean Association sowing guide (Australian Mungbean Association n.d.), mungbeans should not be sown later than January in the Darling Downs region of southern Queensland, the region where most of trials were undertaken. If the planting guide is adhered to, powdery mildew may present later in the crop cycle and yields may be at a lower risk from powdery mildew, reducing the requirement for fungicide applications. An example of this is the 2013 trials, Premer (AM1303), Goolhi (AM1305) Marys Mount (AM1304) and Millmerran (BB1305) where PM manifested 60 days after sowing or later (Table 1), approximately when the plant would be expected to filling pods. The trial reports indicated significant differences in powdery mildew severity between the fungicide treatments, however none of the trials showed a significant difference in grain yield between the same treatments (Northern Growers Alliance 2013c, 2013a, 2013d, 2013b). Crop age may also explain differences between the PM severity and yield analyses reported in this study. Therefore, in lieu of further research, discretion is still required to determine if fungicide applications are

necessary, and consideration given to the crop growth stage and forecast weather conditions at the first sign of PM.

Early detection of PM is vital for an effective fungicidal control response. Data from the trials included in this experiment show regardless of crop sowing date, PM establishes on average in the month of March (Fig. 1), which marks the beginning of autumn in Australia. Such information, together with additional observations, may permit models to predict the likely onset of the disease given weather parameters, providing an early warning system for growers to scout their crop for signs of powdery mildew. The results herein suggest that weekly inspections commencing in March would be ideal to ensure timely fungicide applications at first sign of PM. Scouting at intervals greater than 12–13 days might risk crop losses if a spray schedule does not commence within the aforementioned spray window. More work is needed to understand when “Late” fungicide applications are too late.

We estimate the costs of applying tebuconazole fungicide at \$ 4.18 AU ha⁻¹, if fungicide is purchased at \$ 15 / L (Simpfendorfer and Taylor 2011) and application cost are \$ 2 per hectare (Queensland Government 2019). This meta-analysis estimates average yield savings of between 137.8 kg ha⁻¹ (SE = 33.9) (*Recommended*) and 189.1 kg ha⁻¹ (SE = 53.0) (*Recommended+*). Assuming mungbean is sold at \$ 1200 t⁻¹, these yield savings would save between \$ 71.46 - 234.18 AU ha⁻¹ for a single application at first sign and \$ 83.0 - 337.4 AU ha⁻¹ if a follow-up application is made. Such profit levels, and efficacy for controlling

visual signs of disease, may tempt growers into applying fungicide to control diseases such as powdery mildew when they are unnecessary or unlikely to result in a financially beneficial yield saving. Excessive use of fungicides is likely to lead ongoing problems such as fungicide resistance. The evolution of fungicide resistance can occur quickly, especially when spray schedules are not designed with potential fungicide resistance in mind. Powdery mildew is one such group of fungal pathogens with an extensive track record of rapidly developing fungicide resistance. *P. xanthii* is categorised by the Australian Fungicide Resistance Action Committee (FRAC) as a “High Risk” candidate to evolve fungicide resistance (FRAC 2019; Peterson 1973; Mcgrath 2001; Chin et al. 2001).

To extend the time that a fungicide remains effective against the target pathogen, the strategic use of the fungicide must be considered. To aid in making these decisions and devising a good strategy, DSS tools such as the PowderyMildewMBM consider crop growth stage and weather, to assist decision making whether to spray or not to spray (Diggle 2019) and can help users make an informed decision. Using these sorts of tools can reduce unnecessary and unprofitable fungicide applications, which may help lead to the development of fungicide resistance. To offer more benefit for future research, experiments should record physiological maturity of the crop throughout the season so “crop age” might be considered as a covariate in future analyses.

Currently, the recommendations that are advised to mungbean growers appear to be the best practices for reducing losses due to powdery mildew. Care needs to be taken to

adhere to them, especially avoiding prophylactic sprays, to limit the development of fungicide resistance in the powdery mildew pathogen populations. This meta-analysis provides some direction for future powdery mildew control research in Australian mungbean. However, it does not provide certainty as to when *Late* fungicide applications would be too late to protect yield losses due to the disease. Given the high value of mungbean at the farm-gate, further work could be done in this area that would benefit the Australian mungbean industry.

Supplemental Material

Supplementary material can be found in the following research compendium.

Data Availability

All data and code used in the preparation of this manuscript can be found in the associated research compendium.

Conflict of Interest Statement

The authors of this paper have no conflicting interest in the research presented in this manuscript.

Acknowledgments

The authors wish to acknowledge the work done by Professor Malcolm Riley and Dr Sue Thompson who undertook many of the trials incorporated in this analysis from 2004 - 2015. The Queensland Department of Agriculture, Fisheries and Forestry (DAF). The Northern Grower Alliance and Lawrie Price for contributing data. The Grains Research and Development Corporation who funded this research. Emerson M. Del Ponte was supported by the National Council for Scientific and Technological Development (CNPq) through a Productivity Research Fellowship (PQ) project 310208/2019-0. Thank you to Professor Levente Kiss for his kind assistance in improving this article.

Literature cited

- APVMA. 2014. *Permit to allow minor use of a registered AgVet chemical product for control of powdery mildew in mungbeans*. Australian Pesticides and Veterinary Medicines Authority.
- Australian Mungbean Association. About Us. Available at: <http://www.mungbean.org.au/about-us.html>.
- Australian Mungbean Association. 2016. *Best Management Guide*. Available at: <http://www.mungbean.org.au/best-management-guide.html> [Accessed August 7, 2020].
- Australian Mungbean Association. *Planting mungbeans*. Available at: <http://www.mungbean.org.au/planting.html> [Accessed July 16, 2020b].
- Chankaew, S., Somta, P., Isemura, T., Tomooka, N., Kaga, A., Vaughan, D. A., et al. 2013. Quantitative trait locus mapping reveals conservation of major and minor loci for powdery mildew resistance in four sources of resistance in mungbean [*Vigna radiata* (L.) Wilczek]. *Molecular Breeding*. 32:121–130.
- Chauhan, Y., and Williams, R. 2018. Physiological and Agronomic Strategies to Increase Mungbean Yield in Climatically Variable Environments of Northern Australia. *Agronomy*. 8:83–83.

- Chin, K. M., Chavaillaz, D., Kaesbohrer, M., Staub, T., and Felsenstein, F. G. 2001. Characterizing resistance risk of *Erysiphe graminis* f.sp. *tritici* to strobilurins. Crop Protection. 20:87–96.
- Clarry, S. 2016. The rise and rise of mungbeans. Ground Cover. Available at: <https://grdc.com.au/resources-and-publications/groundcover/ground-cover-supplements/ground-cover-issue-125-pulse-breeding-advances/the-rise-and-rise-of-mungbeans>.
- Conde, B. D., and Diatloff, A. 1991. Diseases of mungbeans. In *Mungbean: the Australian experience. Proceedings of the first Australian mungbean workshop*, St Lucia, Qld. 1991: CSIRO Division of Tropical Crops & Pastures, p. 73–77. Available at: <http://worldveg.tind.io/record/15587>.
- Diggle, A. 2019. *PowderyMildewMBM - Powdery mildew management app for mungbean*. Available at: <https://www.agric.wa.gov.au/apps/powderymildew-mbm-powdery-mildew-management-app-mungbean> [Accessed May 11, 2020].
- Douglas, C. 2008. *National Mungbean Improvement Program DAQ00060 Project Details Project Code: DAQ00060 Project Title: National Mungbean Improvement Program*. Available at: http://era.daf.qld.gov.au/id/eprint/2990/1/GRDC_final_report_DAQ00060.pdf.
- Douglas, C., and McIntosh, P. 2020. Opal-AU: High yielding variety with improved foliar disease protection.
- FRAC. 2019. *Pathogen Risk List*. Available at: <https://www.frac.info/docs/default-source/publications/pathogen-risk/frac-pathogen-list-2019.pdf>.
- Fuhlbohmer, M. J., Ryley, M. J., and Aitken, E. A. B. 2013. Infection of mungbean seed by *Macrophomina phaseolina* is more likely to result from localized pod infection than from systemic plant infection. Plant Pathology. 62:1271–1284.
- Fuhlbohmer, M. J., Ryley, M. J., and Aitken, E. A. B. 1996. *Macrophomina phaseolina* causing leaf spot of mungbean. Australasian Plant Pathology. 25:247–248.
- Humphry, M. E., Magner, T., McIntyre, C. L., Aitken, E. A. b, and Liu, C. J. 2003. Identification of a major locus conferring resistance to powdery mildew (*Erysiphe polygoni* DC) in mungbean (*Vigna radiata* L. Wilczek) by QTL analysis. Genome. 46:738–744.
- Kelly, L. A., Vaghefi, N., Bransgrove, K., Fechner, N. A., Stuart, K., Pandey, A. K., et al. 2021. One crop disease, how many pathogens? *Podosphaera xanthii* and *Erysiphe vignae* sp. nov. identified as the two species that cause powdery mildew of mungbean (*Vigna radiata*) and black gram (*V. mungo*) in Australia. Phytopathology. Available at: <https://apsjournals.apsnet.org/doi/10.1094/PHYTO-12-20-0554-R> [Accessed February 1, 2021].
- Kelly, L., White, J., Sharman, M., Brier, H., Williams, L., Grams, R., et al. 2017. Mungbean and sorghum disease update - GRDC. GRDC Update Papers. Available at: <https://grdc.com.au/resources-and-publications/grdc-update-papers/tab-content/grdc-update-papers/2017/07/mungbean-and-sorghum-disease-update>.

- 400 Lambrides, C. J., and Godwin, I. D. 2007. Mungbean. In *Pulses, Sugar and Tuber Crops*,
401 Springer, p. 69–90.
- 402 Madden, L. V., and Paul, P. A. 2011. Meta-analysis for evidence synthesis in plant pathology:
403 An overview. In *Phytopathology*, The American Phytopathological Society, p. 16–30.
404 Available at: <https://apsjournals.apsnet.org/doi/abs/10.1094/PHTO-03-10-0069>.
- 405 Madden, L. V., Piepho, H. P., and Paul, P. A. 2016. Statistical models and methods for
406 network meta-analysis. *Phytopathology*. 106:792–806.
- 407 Mcgrath, M. T. 2001. *Fungicide Resistance in Cucurbit Powdery Mildew: Experiences and*
408 *Challenges*.
- 409 Ngugi, H. K., Lehman, B. L., and Madden, L. V. 2011. Multiple Treatment Meta-Analysis of
410 Products Evaluated for Control of Fire Blight in the Eastern United States. *Phytopathology*.
411 101:512–522.
- 412 Noble, T. J., Young, A. J., Douglas, C. A., Williams, B., and Mundree, S. 2019. Diagnosis and
413 management of halo blight in Australian mungbeans: a review. *Crop and Pasture Science*.
414 70:195–203.
- 415 Northern Growers Alliance. 2013a. Powdery Mildew Mungbeans - 2013 Goolhi NSW. Farm
416 trials. Available at: <https://www.farmtrials.com.au/trial/20927>.
- 417 Northern Growers Alliance. 2013b. Powdery Mildew Mungbeans - 2013 Mary's Mount
418 NSW. Farm trials. Available at: <https://www.farmtrials.com.au/trial/20926>.
- 419 Northern Growers Alliance. 2013c. Powdery Mildew Mungbeans - 2013 Millmerran QLD.
420 Farm Trials. Available at: <https://www.farmtrials.com.au/trial/20936>.
- 421 Northern Growers Alliance. 2013d. Powdery Mildew Mungbeans - 2013 Premier NSW. Farm
422 Trials. Available at: <https://www.farmtrials.com.au/trial/20925>.
- 423 Pandey, A. K., Burlakoti, R. R., Kenyon, L., and Nair, R. M. 2018. Perspectives and Challenges
424 for Sustainable Management of Fungal Diseases of Mungbean [*Vigna radiata* (L.) R. Wilczek
425 var. *radiata*]: A Review. *Frontiers in Environmental Science*. 6:53–53.
- 426 Paul, P. A., Lipps, P. E., Hershman, D. E., McMullen, M. P., Draper, M. A., and Madden, L. V.
427 2008. Efficacy of Triazole-Based Fungicides for Fusarium Head Blight and Deoxynivalenol
428 Control in Wheat: A Multivariate Meta-Analysis. *Phytopathology*. 98:999–1011.
- 429 Pérez-García, A., Romero, D., Fernández-Ortuño, D., López-Ruiz, F., De Vicente, A., and
430 Torés, J. A. 2009. The powdery mildew fungus *Podosphaera fusca* (synonym *Podosphaera*
431 *xanthii*), a constant threat to cucurbits. *Molecular Plant Pathology*. 10:153–160.
- 432 Peterson, R. A. 1973. Field resistance to benomyl in cucurbit powdery mildew. *Australian*
433 *Plant Pathology Society Newsletter*. 2:27–28.
- 434 Queensland Government. 2019. Mungbean (Irrigated) Darling Downs 2019. AgMargins.
435 Available at: [https://agmargins.net.au/Reports/Details/fced3a80-a762-481c-b812-](https://agmargins.net.au/Reports/Details/fced3a80-a762-481c-b812-9953e33410e6)
436 [9953e33410e6](https://agmargins.net.au/Reports/Details/fced3a80-a762-481c-b812-9953e33410e6).
- 437 R Core Team. 2020. *R: A Language and Environment for Statistical Computing*. Vienna,
438 Austria: R Foundation for Statistical Computing. Available at: <https://www.r-project.org/>.

- 439 Simpfendorfer, S., and Taylor, Z. 2011. *Fungicide Management of Stripe Rust in Wheat: up-*
440 *front vs in-crop options in 2011.*
- 441 Sparks, A., and Kelly, L. 2017. *Mungbean powdery mildew management with fungicide.*
442 Available at: [https://communities.grdc.com.au/field-crop-diseases/mungbean-powdery-](https://communities.grdc.com.au/field-crop-diseases/mungbean-powdery-mildew-fungicide/)
443 *mildew-fungicide/* [Accessed March 1, 2020].
- 444 Thakur, M. P., and Agrawal, K. C. 1995. Epidemiological studies on powdery mildew of
445 mungbean and urdbean. *International Journal of Pest Management.* 41:146–153.
- 446 Thomas, Robertson, M. J., Fukai, S., and Peoples, M. B. 2004. The effect of timing and
447 severity of water deficit on growth, development, yield accumulation and nitrogen fixation
448 of mungbean. *Field Crops Research.* 86:67–80.
- 449 Thompson, S. 2016. Mungbeans vs fungus: two sprays for optimum control - Grains
450 Research and Development Corporation. Ground Cover. Available at:
451 [https://grdc.com.au/resources-and-publications/groundcover/ground-cover-issue-124-](https://grdc.com.au/resources-and-publications/groundcover/ground-cover-issue-124-septemberoctober-2016/mungbeans-vs-fungus-two-sprays-for-optimum-control)
452 *septemberoctober-2016/mungbeans-vs-fungus-two-sprays-for-optimum-control.*
- 453 Viechtbauer, W. 2010. Conducting Meta-Analyses in R with the metafor Package. *Journal of*
454 *Statistical Software.* 36:1–48.
- 455 Wilson, D., Blanche, K. R., and Gibb, K. S. 2001. Phytoplasmas and disease symptoms of
456 crops and weeds in the semi-arid tropics of the Northern Territory, Australia. *Australasian*
457 *Plant Pathology.* 30:159–163.
- 458 Wood, B. A., and Easdown, W. J. 1990. A new bacterial disease of mung bean and cowpea for
459 Australia. *Australasian Plant Pathology.* 19:16–21.
- 460 Zhang, M. C., Wang, D. M., Zheng, Z., Humphry, M., and Liu, C. J. 2008. Development of PCR-
461 based markers for a major locus conferring powdery mildew resistance in mungbean
462 (*Vigna radiata*). *Plant Breeding.* 127:429–432.

463

464 Tables and Figures

465 Table 1: Mean yield and severity for trials included in the meta-analysis. First sign of
 466 powdery mildew date, number of treatments per trial (n), PM median plot severity of the
 467 non-fungicide treated plots and the severity range in fungicide treatments, bracketed
 468 numbers refer to the number of pooled control treatments summarised; Mean yield in non-
 469 fungicide treated control and the range of yields in the fungicide treatments in tonnes per
 470 hectare.

Trial code	Location	Year	Sowing date	PM onset	n	Control median PM severity (n)	PM severity range	Control mean yield	Yield range (t / ha)
1011/01	Hermitage	2011	2011-01-24	2011-03-28	4	4.83 (2)	2 - 2.33	1.525	1.544 - 1.769
1011/02	Kingaroy	2011	2011-02-02	2011-03-22	6	7.83 (2)	3.33 - 5.33	0.798	0.73 - 1.059
1112/01 ^b	Gatton	2012	2012-02-20	2012-04-02	7	7.5 (1)	2 - 6.3	0.738	0.81 - 0.948
1112/02	Kingaroy	2012	2012-02-03	2012-03-12	7	8 (1)	1.7 - 8	0.751	1.036 - 1.323
AM1303	Premier	2013	2012-12-28	2013-02-28	4	8 (1)	2 - 7.83	1.422	1.334 - 1.45
AM1304	Marys Mount	2013	2012-12-24	2013-03-16	2	3 (1)	3.17 - 3.17	1.094	1.265 - 1.265
AM1305	Goolhi	2013	2013-01-23	2013-03-25	4	7 (1)	1 - 3.75	0.694	0.604 - 0.722
BB1305	Millmerran	2013	2013-01-12	2013-03-13	4	8 (1)	1.62 - 8	0.802	0.744 - 0.803
1415/01	Hermitage	2015	2015-01-19	2015-03-16	5	7.8 (1)	5.4 - 6.6	2.045	2.018 - 2.176
1516/01 ^a	Hermitage	2016	2016-02-03	2016-03-08	7	8 (1)	3 - 7.5	1.803	2.141 - 2.37
1516/02 ^a	Kingaroy	2016	2016-02-11	2016-03-09	7	8.25 (1)	2.25 - 7.75	0.811	0.873 - 1.034
1617/01	Hermitage	2017	2017-02-13	2017-03-24	39	8 (9)	5.33 - 8	0.476	0.327 - 0.683
1617/02	Missen Flats	2017	2017-01-27	2017-03-07	39	9 (9)	7.67 - 9	1.561	1.13 - 2.526
1718/01	Wellcamp	2018	2018-02-13	2018-03-21	18	8.17 (6)	5.67 - 7.67	1.194	1.084 - 1.69
1819/01	Hermitage	2019	2018-02-04	2018-04-12	4	7.5 (1)	5 - 7.17	0.587	0.543 - 0.571
1819/02	Hermitage	2019	2018-02-18	2018-04-12	4	7.83 (1)	4.6 - 7.33	0.289	0.276 - 0.312

471 ^a Trials which did not report powdery mildew (PM) severity variance and were therefore not included in the yield protection meta-
 472 analysis; ^b Trials which reported PM severity variance and not yield variance

473

474 Table 2: Estimated powdery mildew severity mean difference (μ) and percentage powdery
 475 mildew (PM) compared to the no spray control (intercept). Estimates were calculated from
 476 a network meta-analysis of data obtained from grey literature reports of (k) field trials

undertaken in Eastern Australia. P values indicate statistical significance in comparison to the intercept.

<i>Moderator</i>	<i>N^a</i>	<i>k^b</i>	Effect Size					PM control (%)		
			μ^c	<i>se</i>	<i>CI_L^d</i>	<i>CI_U^e</i>	<i>P^f</i>	<i>Efficacy^g</i>	<i>CI_L^d</i>	<i>CI_U^e</i>
Early	13	3	-0.7874	0.1941	-1.1678	-0.4069	< 0.0001	10.43	5.39	15.48
Recommended	28	12	-1.1094	0.2625	-1.6238	-0.5950	< 0.0001	14.70	7.88	21.52
Recommended+	41	11	-3.3270	0.5144	-4.3351	-2.3188	< 0.0001	44.09	30.73	57.45
Late	15	5	-0.6906	0.1154	-0.9167	-0.4644	< 0.0001	9.15	6.15	12.15
Late+	13	2	-1.2617	0.2362	-1.7246	-0.7987	< 0.0001	16.72	10.59	22.86

^anumber of treatment means categorised to each spray schedule; ^bnumber of trials with the respective spray schedule;

^cestimated mean yield determined by the meta-analysis; ^dLower range of the 95% confidence interval; ^eUpper range of the 95% confidence interval; ^findicates the significance between each respective spray schedule and the no spray control (intercept);

^gFungicide spray schedule efficacy; ^dLower range of the 95% confidence interval; ^eUpper range of the 95% confidence interval

Table 3: Estimated mungbean yield mean difference to the no spray control (intercept) for each spray schedule treatment. Yield estimates (μ) were calculated from a network meta-analysis of data obtained from grey literature reports of 'k' field trials undertaken in Eastern Australia. P values indicate statistical significance in comparison to the intercept.

<i>Moderator</i>	<i>N^a</i>	<i>k^b</i>	μ^c	<i>se</i>	<i>CI_{L}</i> ^d	<i>CI_{U}</i> ^e	<i>P^f</i>
<i>Early</i>	13	3	0.0029	0.0439	-0.0830	0.0889	0.9464
<i>Recommended</i>	27	13	0.1378	0.0339	0.0713	0.2043	< 0.0001
<i>Recommended+</i>	40	12	0.1891	0.0530	0.0853	0.2930	0.0004
<i>Late</i>	17	7	0.1374	0.0440	0.0512	0.2237	0.0018
<i>Late+</i>	19	4	0.1301	0.0578	0.0168	0.2433	0.0244

^anumber of treatment means categorised to each spray schedule; ^bnumber of trials with the respective spray schedule; ^cestimated mean yield determined by the meta-analysis; ^dLower range of the 95% confidence interval; ^eUpper range of the 95% confidence interval; ^findicates the significance between each respective spray schedule and the no spray control (intercept)

Figure 1: Box plots of the unsprayed control treatments means for both PM severity (top) and crop yield (bottom), indicating within-season variation (A), and trial treatment means for each spray schedule (B). The middle box plot line indicates the median, box boundaries indicate the lower and upper quartiles; circles indicate trial treatment means for each respective crop season and spray schedule.

Data and code for reproducibility

https://github.com/openplantpathology/Mungbean_PM