

1 **Valorization of waste by-products of rose oil production as feedstuff phytonutrients**

2
3 Stefan G. Dragoev^{a,*}, Dessislava B. Vlahova-Vangelova^a, Dessislav K. Balev^a, Dimitar G.
4 Bozhilov^b, Soleya Z. Dagnon^b

5
6 ^a *Department of Meat and Fish Technology, Technological Faculty, University of Food Technologies,*
7 *Plovdiv, Bulgaria*

8 ^b *Department of Organic Chemistry, Faculty of Chemistry, University of Plovdiv "Paisii Hilendarski",*
9 *Plovdiv, Bulgaria*

10
11 * Corresponding author. Tel: +359 899-829-920; Fax: +359-32-603-005; EM:
12 logos2000lt@gmail.com

14 **Highlights**

- 15 • Identified 30 polyphenol antioxidant components in dry rose petals.
- 16 • Found 6 new glycosides of gallic acid except glycosides of kaempferol and quercetin.
- 17 • Supplementation of 545 mg DDRP/kg/d can be successfully used in pig's husbandry.
- 18 • This DDRP supplementation isn't effective for growth performance of broilers or lambs.

20 **Abstract**

21 The aim of this study was to evaluate possibility for valorization of the dry pressed distilled
22 rose petals (DDRP) as a feedstuff phytonutrients in animal husbandry. In connection with the
23 chemical composition and radical scavenging activity of polyphenol complex in rose waste
24 products were studied. The polyphenol composition in dry rose petals, DDRP and waste water
25 after distillation was identified and quantified. The chromatographic analysis was conducted by
26 HPLC-PDA and LC-MS. The experiments were conducted with bird, monogastric and ruminants
27 representatives. A total of 40 one days old chicks from the hybrid combination Ross 308 were
28 distributed into two groups each containing of 20 birds fed for 49 days. The control group C fed
29 with basal diet vs. experimental group R fed with the same fodder plus 40 mg DDRP/kg/d. A
30 total of 72 Danube White 155 days old fattening pigs (both sexes - 36 male and 36 female) of

31 72.500 ± 1.937 kg were distributed into three groups (one control C and two experimental R1
32 and R2 supplemented with 0.255 g DDRP/kg/d or 0.545 g DDRP/kg/d). Each one group (24
33 piglets) was fed for 45 days. A total of 30 numbers of 65 days old lambs from the Synthetic
34 population of Bulgarian milk sheep breed lambs (both sexes - 15 male and 15 female) with
35 average life weight 20.500 ± 1.039 kg were divided into two groups (control C and experimental
36 R) each one containing of 10 animals fed for 50 days. The experimental group was fed with
37 combined feed for lambs granules and ground alfalfa supplemented either 0.545 g DDRP/kg/d.
38 Animals were assessed for average daily weight gain and feed conversion ratio. Thirteen
39 glycosides of kaempferol, ten glycosides of quercetin, six glycosides of gallic acid and the two
40 flavonol aglycones have been identified in dry rose petals. Those polyphenols possess high
41 antioxidant activity and positively influence the growth performance of pigs. The
42 supplementation of pig's feed with 545 mg DDRP/kg/d increased ($p < 0.05$) with 6.73% the total
43 feed consumption and with 27.05% the average daily weight gain, and decreases with
44 approximately 16% the feed conversion ratio comparing to control group (C) pigs fattening
45 without DDRP supplementation. The studied concentrations of DDRP were not effective in
46 small ruminants and poultry husbandry.

47 *Keywords:* dry pressed distilled rose petals; polyphenols; feedstuff supplements; livestock;
48 growth performance

49 *Abbreviations:* ADG, average daily weight gain; AFC, average feed consumption; DDRP, dry
50 pressed distilled rose petals; DPPH•, 2,2-diphenyl-1-picrylhydrazyl; FCR, feed conversion ratio;
51 FRAP, ferric ion reducing antioxidant power; FWHM, full width at half maximum; HCD,
52 higher-energy C-trap dissociation; HPLC, high-performance liquid chromatography; HPLC-
53 PDA, high-performance liquid chromatography-photodiode array; HRMS, high-resolution mass

54 spectrometer; GAE, gallic acid equivalents; LC-MS, Liquid chromatography–mass spectrometry;
55 PDA, photodiode array; NCE, normalized collision energy; SD, standard deviation; TE, Trolox
56 equivalents; UHPLC, ultrahigh- performance liquid chromatography; UHPLC-MS/MS, ultra-
57 high-performance liquid chromatography-tandem mass spectrometry.

58 **1. Introduction**

59 Oilseed rose (*Rosa damascena* Mill.) processing is widely distributed on the Balkan
60 Peninsula countries Bulgaria (Rusanov et al., 2014) and Turkey (Erbas and Baydar, 2016). The
61 rose oil and rose water are highly valued in perfumery and cosmetics (Aggarwal and Kaur,
62 2017). Recently Özkan et al. (2004) have been reported the strong antioxidant capacity and
63 antibacterial activity of *Rosa damascena* fresh and spent flower extracts against 15 pathogenic
64 bacterial species. The data of Baydar and Baydar (2013) confirm the high antiradical activity and
65 antioxidant capacity of several phenolic compounds discovered in hot and cold methanolic
66 extracts of oil-bearing rose (*Rosa damascena* Mill.). They suppose that oil-bearing rose by-
67 products and wastes can be discussed as natural antioxidant sources. In the literature can be
68 found only few publications discussing the optimisation of technology and the extraction
69 procedure of rose by-products and waste with the purpose of their utilization. An original
70 approach for hydrothermal gasification of *Rosa damascena* residues was published by Akgül et
71 al. (2014). They concluded that the rose wastes have potential to be a future source for hydrogen
72 production. It has been offered industrial wastes of *Rosa damascena* to be regarded as a source
73 of water-soluble pectic extracts (Slavov et al., 2016). In connection with this a method for
74 recovery of biologically active substances from rose (*Rosa damascena* Mill.) by-products was
75 proposed (Slavov et al., 2017).

76 [Schieber et al., \(2005\)](#) first have analyzed the flavonol glycosides extracted from distilled
77 rose petals of oil-bearing rose. Twenty two major compounds were identified including
78 kaempferol and quercetin glycosides, quercetin 3-O-galactoside and quercetin 3-O-xyloside. It
79 has been shown that the kaempferol glycosides comprise about 80% of the quantified
80 compounds. By LC-MS analysis were indentified thirteen kaempferol and eleven quercetin
81 glycosides in 30% v/v water-ethanolic extract from industrially distilled rose petals ([Shikov et
82 al., 2008](#)).

83 The by-products of Taif rose (*Rosa damascena trigintipetala* Dieck) have been shown to be
84 a source of natural antioxidants with strong antioxidant activity ([Abdel-Hameed et al., 2012](#)) too.
85 Their free radical scavenging potential correlated with the biochemical components content in
86 red rose petals ([Pal et al., 2018](#)). [Rusanov et al. \(2014\)](#) have reported the presence of flavan-3-
87 ols, flavanones, flavonols and flavones in rose oil distillation wastewater. Due to their
88 pronounced antioxidant and antibacterial properties, rose by-products or wastes have found many
89 applications in various branches of the food industry ([Slavov et al., 2017](#)). They were suggested
90 as colour stabilizers of strawberry beverage ([Mollov et al., 2007](#)) and texture-improved canned
91 strawberries ([Shikov et al., 2012](#)), and as natural antioxidants added to meat and sausages
92 ([Oswell et al., 2018](#)). The potential of the rose oil industry wastes was discussed to find
93 application in liqueurs preparations ([Vasileva et al., 2019](#)) and in probiotic lactic acid bacteria
94 dairy products ([Dimitrova et al., 2019](#)). The dry distilled rose (*Rosa damascena* Mill.) petals
95 were used for enrichment of broiler's feed ([Balev et al., 2015](#)). The utility model relating to the
96 composition of the feed supplement for livestock and poultry, based on dry distilled rose petals
97 was registred, as well ([Vlahova-Vangelova et al., 2018](#)).

98 When consumed regularly by humans, polyphenols have been associated with a reduction in
99 the incidence of diseases such as cancer, obesity, diabetes and heart disease (Koch, 2019). The
100 ability of these natural antioxidants, to scavenge several oxygen and nitrogen free radicals has
101 been associated to the health benefits of diets rich in polyphenols (Abdel-Hameed et al., 2012;
102 Baydar and Baydar, 2013; Pal et al.,2018). From this point of view the objectives of this research
103 were to determine the possibilities for dry pressed distilled rose petals (DDRP) valorization as a
104 feedstuff phytonutrients in animal husbandory, to study the chemical composition and radical
105 scavenging activity of polyphenol complex in rose (*Rosa damascena* Mill.) waste products, to
106 identify and quantify the polyphenol composition in dry rose petals, dry pressed distilled rose
107 petals and waste water (liquid aqueous phase after distillation).

108 **2. Materials and methods**

109 *2.1. Rosa damascena* Mill. Sampling

110 Dry rose petals and distilled waste by-products such as dry pressed distilled rose petals
111 (DDRP) and waste water (liquid aqueous phase after distillation) were analyzed. The waste by-
112 product was collected after distillation column of the installation of rose oil extraction by the
113 company Bulattars Productions (Skobelevo village, Bulgaria). It was compressed for 12 h at
114 room temperature at a pressure of 303.975 KPa. The obtained presses were dried with hot air
115 (60°C, 6 h) to equilibrium humidity. The dry matter in dry distilled rose petals was 97.92 ± 0.19
116 g/100 g and 97.98 ± 0.21 g/100 g in dry pressed distilled rose petals, respectively. The dry
117 residues were ground in a ball mill.

118 The liquid aqueous phase from the rose petals distillation was collected after the distillation
119 column of the rose oil extraction installation. It was concentrated by evaporation during the

120 boiling in open pots. A concentrate was obtained from the liquid aqueous phase of the distilled
121 rose petals.

122 The distilled rose petals were dehydrated by presses and then was dried. The distilled liquid
123 phase was concentrated by evaporation of the water contained therein.

124 *2.2. Experimental procedures*

125 *2.2.1. Polyphenol indices and antioxidant activity*

126 *2.2.1.1. Determination of total anthocyanins*

127 The amount of total monomer anthocyanins was determined by the pH differential method
128 ([Giusti and Jing, 2007](#)) based on the property of anthocyanin pigments to change the color with
129 pH.

130 *2.2.1.2. Determination of total polyphenols*

131 The content of total polyphenols was determined by the method of [Singleton and Rossi](#)
132 ([1965](#)). The results obtained are presented as gallic acid equivalents (GAE).

133 *2.2.1.3. Determination of antioxidant activity using DPPH• free radical*

134 The DPPH• radical scavenging ability was determined by the method of [Brand-Williams et](#)
135 [al. \(1995\)](#). The results obtained are presented as Trolox (TE) equivalents.

136 *2.2.1.4. Determination of ferric ion reducing antioxidant power (FRAP-test)*

137 The FRAP was determined by the method of [Benzie and Strain \(1996\)](#). The results are
138 presented as Trolox (TE) equivalents.

139 *2.3. Chromatographic analysis*

140 *2.3.1. Ultrasound-assisted extraction of polyphenols*

141 0.2 g powder of dry rose petals and dry pressed distilled rose petals samples were weighed.
142 Five instances of every sample were prepared. The waste rose water was injected directly in the

143 HPLC device after filtration. The polyphenols were analyzed in their glycoside form (and
144 therefore no hydrolyzed plant extracts were prepared) by extracting the samples with 10 ml 70%
145 (v/v) aqueous methanol in an ultrasound bath for 40 min at room temperature (25°C). The
146 extracts were filtrated under reduced pressure. The volume of the samples was adjusted to 10 ml
147 and passed through a membrane filter 0.45 μm prior to HPLC analysis.

148 2.3.2. HPLC-PDA analysis of polyphenols

149 The instrumentation used for HPLC analysis consisted of quaternary mixer Smartline
150 Manager 5000, pump Smartline 1000 and PDA 2800 detector (Knauer, Germany). Separation of
151 polyphenol components was performed on Kromasil C18, 15 cm \times 4.6 mm i.d. 5 μm particle size
152 (Supelco, USA. The chromatography was carried out using as mobile phase A mixture from 95
153 parts 2% formic acid in water and 5 parts mobile phase B. As solvent B was used mixture from
154 10 parts 2% formic acid in water and 90 parts 2% formic acid in acetonitrile. The polyphenols
155 were eluted with a gradient system as follow: 0-15 min, 100 % - 90 % A (0 - 10 % B), 15 - 25
156 min, 80 % A (20 % B), 25 - 55 min, 50 % A (50 % B), 55 - 60 min, 0 % A (100 % B).

157 Mobile phase flow rate was set by 1.0 ml/min; sample volume was 20 μl . The polyphenols
158 were monitored at 320 nm, 340 nm, 352 nm and 280 nm.

159 The spectral characteristics of eluting peaks of each sample, scanned with a PDA detector
160 ($\lambda=200\text{--}400$ nm), were compared with those of authentic standards, gallic acid, rutin, quercetin-
161 3-O-glucoside, quercetin-3-O-galactoside, quercetin and kaempferol (Sigma Aldrich). The
162 identification of compounds was made by summarizing the data for retention times, UV and MS
163 spectra of standards and the peaks in the samples, and previously published information. The
164 MS/MS data for the molecular mass and the fragmentation of deprotonated molecular ions [M-
165 H]⁻ were worked up by assigning the structure of compounds.

166 Quantification of main polyphenol components was performed by using the data from the
167 fingerprint profiles obtained from HPLC-PDA analysis at 280nm and 352nm.

168 The calibration curves were prepared from stock solutions of analytical standards at a
169 concentration of 1000 mg.L⁻¹ in methanol by successive dilution until the optimal range of
170 application for each compound. The calibration standards and the samples were injected in
171 duplicate.

172 2.3.3. LC-MS analysis

173 The ground powder of dry rose petals (10 g) was extracted with 500 ml ethylacetate: methanol
174 (1:1 v/v) in an ultrasonic bath for 40 min at room temperature (25°C). The extract was filtered
175 under reduced pressure. The liquid was evaporated in a rotary evaporator at 40°C to give a
176 residue (1.1426 g), which was dissolved in water and successively partitioned among hexane,
177 chloroform and ethyl acetate, each (10 x 30 ml). The obtained fractions were evaporated to
178 dryness. The chloroform and ethyl acetate fractions were dissolved in 4 ml methanol each and
179 were subjected to UHPLC-MS/MS analysis ([Bojilov et al., 2017](#)).

180 The ethyl acetate (23.67%) and chloroform (3.79%) fractions obtained after liquid–liquid
181 extraction of the aqueous solution of dry rose petals crude extract were analyzed. The LC-MS
182 analysis was performed on a Q Exactive Plus high-resolution mass spectrometer (HRMS) with
183 heated electrospray ionization source (HESI-II) (Thermo Fisher Scientific, Inc., Bremen,
184 Germany) equipped with a Dionex Ultimate 3000RSLC ultrahigh-performance liquid
185 chromatography (UHPLC) system (Thermo Fisher Scientific, Inc.). Operating conditions for the
186 HESI source in negative ionization mode were: 2.5 kV spray voltage, 320°C capillary and probe
187 heater temperature, sheath gas flow rate 38 units, auxiliary gas flow 12 units (units refer to
188 arbitrary values set by the Exactive Tune software) and SLens RF level 50.0. Nitrogen was used

189 for sample nebulization and collision gas in the HCD cell. The LC–MS method was operating in
190 full scan-ddMS2/Top5 with the following settings: 70,000 FWHM resolution (at m/z 200), AGC
191 target 3e6, max. IT 100 ms and mass range m/z 100-1500 were chosen, while ddMS2 conditions
192 were set to resolution 17,500 FWHM (at m/z 200), AGC target 1e5, max. IT 50 ms, isolation
193 window 2.0 amu and step normalized collision energy (NCE) was set to 10, 20 and 30. The
194 UHPLC separations were performed on a Kromasil Eternity XT C18, 1.8 μ m, 2.1 \times 100mm
195 column (AkzoNobel, Sweden) with a binary mobile phase consisting of solution A: 0.1% formic
196 acid in water and solution B: 0.1% formic acid in acetonitrile. The following step gradient
197 program was used: 0 min, 95% A; 0.5 min, 95% A, 6 min, 86% A. 12 min, 76% A; 26 min, 48%
198 A; 28 min, 10% A; 30 min, 10% A; 30.5 min, 95% A. Equilibration time was 4.5 min prior to
199 injection, the flow rate was 0.3 mL/min and the sample volume was 1 μ L. The column
200 compartment temperature was set to 40°C. The data acquisition was accomplished with Xcalibur
201 (Thermo Scientific) software ver. 4.0. The calculation of the exact masses and mass
202 measurement errors, prediction of molecular formulas and simulation of monoisotopic profiles
203 were carried out with Xcalibur ver. 4.0 or FreeStyle ver. 1.5 software (Thermo Scientific).

204 *2.4. Design of experiments for valorization of DDRP as phytonutrient*

205 The experiments were conducted in accordance with Art. 14 of Part V. Breeding and
206 Livestock Units from European Convention for the Protection of Vertebrate Animals used for
207 Experimental and Other Scientific Purposes, Commission Recommendation 2007/526/EC and
208 Council Regulation (EC) No 1099/2009. The experiments were approved by the Bulgarian
209 Scientific Ethics Committee and requirements of the Council Directive 2010/63/EC were met.
210 During the fattening period, the animals were reared in accordance with the requirements of
211 Bulgarian Ordinance No 21 of December 14, 2005.

212 2.4.1. Pigs

213 A total of 72 Danube White 155 days old fattening pigs (both sexes - 36 male and 36
214 female) of 72.500 ± 1.937 kg (mean \pm SD) were used. They were housed in barn equipped with
215 individual pens with feeders and drinkers, in facilities at the Experimental farm of Agricultural
216 Institute, Shumen, Bulgaria. During the experiment, the temperature was between 19 to 27°C.
217 Feed was available ad libitum. Water was provided by nipples drinkers ad libitum. The pigs were
218 randomized by origin, age, sex, weight and were distributed into three groups (one control and
219 two experimental), each containing of 24 animals fed for 45 days with two different levels of
220 DDRP supplementations. The control group (C) was fed basal diet (Table 1) supplied by Vasil
221 Kostov feed factory, Lyuben Karavelovo village, Varna district, Bulgaria. The other two
222 experimental groups were fed with the same diets containing either 0.255 g DDRP/kg/d (R1) or
223 0.545 g DDRP/kg/d (R2). Residual feed was monitored daily and was weighed and subtracted
224 from of the daily amount of feed consumed. Individual daily doses of the supplements were
225 calculated according to previous weighing of animals, mixed with feed and given with the
226 morning feeding. Pigs were weighed every two weeks. At the end of the experiment, the age of
227 the pigs was 200 days and the average live weight was at about 110.300 ± 2.274 kg.

228 2.4.2. Lambs

229 A total of 30 numbers of 65 days old lambs from the Synthetic population of Bulgarian milk
230 sheep breed lambs (both sexes - 15 male and 15 female) with average life weight 20.500 ± 1.039
231 kg (mean \pm SD) were used. They were housed in a total indoor barn in facilities at the
232 Experimental Farm of Agricultural Institute, Shumen, Bulgaria. The lambs were divided into
233 three groups (one control and two experimental), each containing of 10 animals fed for 50 days.
234 During the experiment, the temperature was 19 - 25°C. Feeding the lambs was ad libitum in

235 group boxes, with access to water and salt. Feed and water were available. Individual daily doses
236 of the supplements were calculated according to previous weighing of animals, mixed with feed
237 and given with the morning feeding. The control group (C) was fed combined feed for lambs
238 granules and ground alfalfa (grower and finisher basal diets Table 1) supplied by Vasil Kostov
239 feed factory, Lyuben Karavelovo village, Varna district, Bulgaria. The experimental group (R)
240 was fed with the same diet supplemented either 0.545 g DDRP/kg/day. Daily control of the
241 amount of combined feed consumption during the experiment was made. Residual feed was
242 weighed and subtracted from of the daily amount of feed consumed. Lambs were weighed every
243 two weeks. At the end of the experiment, the lambs were 115 days old with the average live
244 weight at about 36.415 ± 0.849 kg.

245 2.4.3. Broilers

246 A total of 40 one days old chicks from the hybrid combination Ross 308 were purchased
247 from the Bovans Bulgaria hatchery, Chirpan, Bulgaria. They were housed in a breeding room at
248 the Experimental farm of Agricultural University, Plovdiv, Bulgaria. During the experiment, the
249 temperature was between 19 to 25°C. Feed and water were available ad libitum. The chicks were
250 distributed into two groups each containing of 20 birds fed for 49 days. The control group (C)
251 was fed basal diet (Table 1). The combined feed was purchased from Viand Ltd, Sofia, Bulgaria.
252 The experimental group (R) was fed with the same fodder containing either 40 mg DDRP/kg/d.
253 A DDRP supplementation was made after its pre-grinding. Individual daily doses of the
254 supplements were calculated according to previous weighing of broilers, mixed with feed and
255 given with the morning feeding. The body weight of broilers was controlled at seven days
256 intervals. Residual feed was monitored daily and was weighed and subtracted from of the daily

257 amount of feed consumed. Feeding and watering the broilers was done ad libitum. At the end of
258 the experiment the live weight of the broilers was at about 3.460 ± 0.089 kg.

259 *2.5. Determination of total feed consumption and growth performance*

260 The total feed consumption and growth performance was determined by weight
261 measurements.

262 *2.6. Feed conversion ratio calculation*

263 The feed conversion ratio (FCR) was calculated as a ratio between the average feed
264 consumption (AFC) and the average daily growth (ADG) of the animals or birds. The

265 FCR was calculated by the formula (1):

$$266 \quad \text{FCR} = \text{AFC}/\text{ADG} \quad (1), \text{ where:}$$

267 AFC - average feed consumption, kg;

268 ADG - average daily growth, kg;

269 *2.7. Statistical Analysis*

270 Five repetitions of every sample were analyzed. Data are expressed as means \pm standard
271 deviation (\pm SD), $n = 5$. Microsoft Excel Office Professional Plus 2010 was used to process the
272 statistics.

273 **3. Results**

274 To achieve the fullest possible utilization of valuable rose biomass our attention was
275 directed to define the possibility for valorization of waste products after isolation of essential oil
276 from *Rosa damascena* as a feedstuff in the animal husbandry.

277 *3.1 Basic polyphenol indices and antioxidant activity of DDRP*

278 Hence, at first a careful study of polyphenol complex of the waste products was
279 undertaken with special attention on dry pressed distilled rose petals (DDRP) regarded as a much

280 suitable feedstuff for animal's feeding. The basic polyphenol indices as the content of total
281 polyphenols and anthocyanins reveal their high amount in the dry pressed distilled rose petals.
282 DDRP contains total polyphenols 7504.00 ± 24.00 mg GAE/100 g and 92.10 ± 0.25 mg
283 anthocyanins/100 g dry matter. This is a reason DDRP to have a strong radical scavenging
284 activity against DPPH• radical ($39138.90 \mu\text{mol TE}/100 \text{ g}$) and to express high ferric ion
285 reducing antioxidant power (FRAP = $35550 \mu\text{mol TE}/100 \text{ g}$).

286 *3.2. Identification and quantification of polyphenol components in dry rose petals and waste* 287 *after distillation of essential oil*

288 On [Figure 1](#) are presented the fingerprint chromatographic profiles of dry rose petals, dry pressed
289 distilled rose petals (DDRP) and waste water at 280nm and 352nm. In the three fingerprint
290 profiles the UV spectra from PDA detector show mostly peaks of gallic acid (2min – 20min),
291 quercetin (24min-26min) and kaempferol (27min-36min) derivatives. The chromatographic
292 profiles of dry rose petals and distilled waste by-products DDRP and waste water contain
293 components with the same spectra, hence no qualitative differences in the polyphenol
294 composition were observed. The careful chromatographic analysis revealed the abundance of
295 polyphenolic compounds in the dry rose petals and DDRP. Different proportions of three main
296 groups of polyphenolic compounds were found in dry rose petals, in dry pressed distilled rose
297 petals and in waste water after isolation of essential oil from *Rosa damascena* ([Table 2](#)). In the
298 waste water was established the lowest amount of all components. The DDRP contains
299 significant amount of glycosides as well as kaempferol ([Table 2](#)). Approximately 50% from all
300 polyphenolic compounds determined in rose petals were found in DDRP. The DDRP was
301 characterized with the highest content of kaempferol. On contrary, the waste water profile was
302 leaking of kaempferol and in general the polyphenol components were in lower amount.

303 The fractionation of dry rose petals led to a small chloroform fraction (3.79 %) in which both
304 flavonol aglycones quercetin and kaempferol were identified. In the ethylacetate fraction
305 (23.67%) only glycosides were ascertained (Figure 2). In summary 30 polyphenol components
306 were identified in dry rose petals as presented in Table 3. Mainly three groups of polyphenolic
307 compounds in following order comprise the polyphenol complex of *Rosa damascena* dry petals:
308 glycosides of kaempferol > glycosides of quercetin > glycosides of gallic acid (Table 3).
309 Thirteen glycosides of kaempferol, 10 glycosides of quercetin, 6 glycosides of gallic acid and the
310 two flavonol aglycones are identified. The MS/MS data reveal the predominant number of
311 kaempferol glycosides in the polyphenol complex of *Rosa damascena* petals. The fragmentation
312 of glycosides by low collision energy is leading to decomposition of the glycosidic bond and loss
313 of 162 Da, 146 Da, 152 Da, 132 Da и 42 Da, which are hexose, (glucose, galactose, rhamnose),
314 galloyl group, pentose and acetyl group. The ESI-MS/MS spectra show the ions of quercetin (m/z
315 301), kaempferol (m/z 285) and gallic acid (m/z 169) (Table 3). Our data indicate the presence of
316 flavonol glycosides mainly kaempferol and quercetin glycosides in the dry rose petals with
317 exception of six glycosides of gallic acid (1-6) and additional isomers of quercetin galloyl
318 hexoside and kaempferol disaccharide, which are identified for the first time. The galloyl
319 glycosides undergo fragmentation with elimination of galloyl group (152 Da) and hexose (162
320 Da). The characteristic ions for gallic acid are m/z 125 and m/z 107 (Table 3).

321 3.3. DDRP as feed supplement

322 Three types of experiments for valorization of DDRP were conducted concerning feeding pigs,
323 broilers and lambs with addition of rose waste to the basal diet. Similar *in vivo* investigations are
324 scarce therefore the results are with high practical value.

325 3.3.1. Pigs

326 It was found the addition of 0.545 g DDRP/kg/d (Experimental group R2) to the forage of
327 fattening pigs contributes for an increasing of the total feed consumption with 6.73% ($p < 0.05$)
328 and of the average daily growth with 27.05%, comparing to control group (C) (Table 4).
329 Therefore, the feed conversion ratio in pigs from experimental group (R2) was with
330 approximately 16% lower than those in pigs from control group (C) (Table 4). The conclusion
331 was made that the supplementation of feed with 0.545 g of DDRP/kg/d can intensify the Danube
332 white pig's growth performance comparing with pigs from control group (C).

333 3.3.2. Chickens

334 The 40 mg DDRP/kg/d supplementation of feedstuffs for broilers has no enough significant
335 effect ($p \geq 0.05$) on the total feed consumption, average daily growth and feed conversion ratio in
336 broilers (Table 4).

337 3.3.3. Lambs

338 Similarly, the 545 mg DDRP/kg/d supplementation of feedstuffs for lambs does not have a
339 effective sufficient ($p \geq 0.05$) impact on their growth performance, the total feed consumption,
340 average daily growth and ultimately on the feed conversion ratio (Table 4).

341 4. Discussion

342 In the last decade the increased essential oil production generates huge amount of waste
343 containing bioactive components. Waste waters and the solid biomass are retained and in most
344 cases simply discarded after the distillation process. Various approaches for rose waste
345 valorization have been proposed during the last years with purpose to utilize the valuable
346 biomass (Slavov et al. 2017). The determined high amount of total polyphenols and anthocyanins
347 in the dry pressed distilled rose petals confirm the results presented by Özkan et al. (2004) in
348 residues of spent flowers of *Rosa damascena* Mill. after steam distillation and by Dina et al.
349 (2018) in the rose hydrodistillation byproducts remaining as aqueous extract. The final extract

350 has a significant antioxidant activity and could be find application on the market of cosmetics,
351 nutraceuticals or phytotherapeutics (Dina et al., 2018).

352 Polyphenols are polar compounds containing more than one hydroxyl group, hence for their
353 extraction 70% methanol is recommended as the most selective and appropriate solvent for
354 HPLC analyses with high precision (Dagnon et al., 2018). Proper identification and
355 quantification of compounds is needed to ascertain the polyphenol composition demanding to
356 develop HPLC-PDA fingerprint chromatographic profiles. For this reason, by the analysis of
357 specific extract, the analysis time, the type of acid in the mobile phase and its concentration, and
358 the slope of the gradient require optimization. The fingerprint profile was accepted when it
359 contained maximum number of good separated peaks (Dagnon et al., 2018). In the case of *Rosa*
360 *damascena* the best separation of both quercetin-3-*O*-galactoside and quercetin-3-*O*-glucoside,
361 which are very similar, was achieved on Kromasil column by the described mobile phase and
362 gradient.

363 Similarly to our results the presence of flavonol glycosides in distilled rose petals has been
364 reported by Schieber et al. (2005). They have identified 22 kaempferol and quercetin glycosides
365 unlike our data that are identified 30 polyphenolic compounds: 13 kaempferol glycosides, 10
366 quercetin glycosides, 6 glycosides of gallic acid and 2 flavonol aglycones. The data in our study
367 are leading to suggest that most of the bioactive flavonoid glycosides are retained unchanged in
368 DDRP after distillation comprising predominantly kaempferol derivatives. Nevertheless the
369 amounts of quercetin and kaempferol glycosides are quite the same.

370 Similarly to us Abdel-Hameed et al. (2012) reported that the residues of Taif rose possess
371 radical scavenging activity, antioxidant capacity and reducing power activity. Abdel-Hameed et

372 al. (2012) confirm the phenolic compounds, especially flavonols are the major antioxidant active
373 components in the Taif rose residues.

374 The found increasing of the total feed consumption the average daily growth and decreasing
375 of feed conversion ratio in pigs supplemented with 0.545 g DDRP/kg/d can be explained with
376 antioxidative and antimicrobial actions of their phytogetic compounds which can promote the
377 animal health (Windisch et al., 2008). As the polyphenols in DDRP probably have ability to
378 stimulate the digestive tract they may be increase the appetite of pigs (Frankič et al. 2009) and
379 can find an application as growth promoters in pigs' production (Valenzuela-Grijalva et al.,
380 2017). Yang et al. (2015) suggested that the established increase in feed palatability could be due
381 to the antioxidative effects of DDRP polyphenols, which might contribute to preserving release
382 of unfavorable odors and a good quality of diets. On the other hand, the investigated
383 phytonutrients exert a specific effect on the gut functions, stabilizing microbial eubiosis (Jamroz
384 et al., 2006). They show an antibacterial activity against 15 bacteria species (Özkan et al., 2004).
385 Thus they improve of nutrient digestion and absorption (Valenzuela-Grijalva et al., 2017). Last
386 but not the least, discussed photogenic compounds may have anti-inflammatory effect too (Pitman
387 and Blumberg, 2000). Superior feed conversion ratios of finisher pigs can also be achieved
388 through not mixing rations on the operation (Losinger, 1998).

389 Our results indicate that the supplementation enrichment of broiler feed with 40 mg
390 DDRP/kg/d is not sufficiently effective to increase the total feed consumption, average daily
391 growth and to decrease the feed conversion ratio. Additional studies are needed for determining
392 the effective doses of DDRP supplementations in broilers because the respective experimental *in*
393 *vivo* evidences are still quite limited (Hashemi and Davoodi, 2011). Breeding and genetics are
394 important factors for 6 weeks of age chickens housed in groups (Leenstra and Pit, 1987). When

395 planning further research it has to pay attention on individual selection for feed conversion.
396 Probably changes in body composition and in growth pattern contribute to the favorable feed
397 conversion ratio from 21 to 42 days of age chickens ([Pasternak and Shalev, 1983](#)). Next
398 important factor we have to pay attention in future research it is heat stress and connected to it
399 body temperature ([Cooper and Washburn, 1998](#)). Those authors shown the individual gain, feed
400 consumption, and feed conversion ratio from 28 to 49 d directly depend on a heat stress
401 environment temperature (32°C). This means that should be avoided to perform the experiments
402 during the summer months, when the air temperature normally reached similar values.

403 Those findings confirm that the addition of 545 mg DDRP/kg/d to lambs' feed did not have
404 a significant effect on the total feed consumption, average daily gain and feed conversion ratio.
405 The explanation of those finding were likely due to the fact that these amounts of DDRP could
406 not improve the condition and feed intake because they are not modulators of ruminal
407 fermentation in small ruminants ([Surai, 2014](#)) and have no anabolic activity on target tissues
408 ([Bahadoran et al., 2013](#)).

409

410 **4. Conclusions**

411 In conclusion the dry distilled rose petals contain three groups of polyphenolic compounds:
412 glycosides of kaempferol > glycosides of quercetin > glycosides of gallic acid. Therefore the
413 DDRP characterizes with significant amount of polyphenols and high antioxidant activity.

414 The DDRP in the investigated concentrations showed limited potential for application as
415 feed supplements in small ruminants and poultry husbandry. On contrary supplementation of
416 feed with 0.545 g of DDRP/kg/d improved Danube white pig's performance.

417

418 **Conflict of interests**

419 The authors declare no conflict of interests.

420

421 **Acknowledgements**

422 The current project was funded by the Bulgarian National Science Fund (BNSF), Ministry of
423 Education and Science of Republic of Bulgaria of state contract No DN 06/8 of December 17,
424 2016 “Study of the mechanism of biological active compounds of plant origin accumulation in
425 the organism of Bulgarian breed agricultural animals and their impact on the meat quality as a
426 natural functional food”. Authors acknowledge the help of Assoc. Prof. Paraskev Nedialkov
427 from Department of Pharmacognosy, Faculty of Pharmacy, Medical University of Sofia, 2
428 Dunav Str. 1000 Sofia, Bulgaria by UHPLC-MS/MS analyses.

429 **References**

- 430 Abdel-Hameed, E.-S., Bazaid, S. A., Shohayeb, M.M. 2012. Total phenolics and antioxidant
431 activity of defatted fresh Taif rose, Saudi Arabia. Br. J. Pharm. Res., 2, 129-140.
432 [https://doi.org/ 10.9734/BJPR/2012/1493](https://doi.org/10.9734/BJPR/2012/1493)
- 433 Aggarwal, P., Kaur, S. 2017. Technology development for the preparation, concentration and
434 utilization of rose extract in different valuable products and by products with retention of
435 color and flavor. Pharma Innov. J., 6, 189-193. [http://www.thepharmajournal.com/
436 archives/2017/vol6issue6/PartC/6-6-9.pdf](http://www.thepharmajournal.com/archives/2017/vol6issue6/PartC/6-6-9.pdf)
- 437 Akgül, G., Madenoğlu, T.G., Cengiz, N.Ü., Gökkaya, D., Sağlam, M., Yüksel, M. 2014.
438 Hydrothermal gasification of Rosa Damascena residues: gaseous and aqueous yields. J.
439 Supercr. Fluids, 85, 135-142. <https://doi.org/10.1016/j.supflu.2013.11.007>

- 440 Bahadoran, Z., Mirmiran, P., Azizi, F. 2013. Dietary polyphenols as potential nutraceuticals in
441 management of diabetes: a review. *J. Diab. Metab. Disor.*, 12, 431-439.
442 <http://www.jdmdonline.com/content/12/1/43>
- 443 Balev, D., Vlahova-Vangelova, D., Mihalev, K., Shikov, V., Dragoev, S., Nikolov, V. 015).
444 Application of natural dietary antioxidants in broiler feeds. *J. Moun. Agric. Balk.*, 18, 224-
445 232. http://www.rimsa.eu/images/stockbreeding_18-2.pdf
- 446 Baydar, N.G., Baydar, H. 2013. Phenolic compounds, antiradical activity and antioxidant
447 capacity of oil-bearing rose (*Rosa damascena* Mill.) extracts. *Ind. Crops Prod.*, 41, 375-380.
448 <https://doi.org/10.1016/j.indcrop.2012.04.045>
- 449 Benzie, I. F., Strain, J. J. 1996. The ferric reducing ability of plasma (FRAP) as a measure of
450 "antioxidant power": the FRAP assay. *Anal. Biochem.*, 239, 70-76.
451 <https://doi.org/10.1006/abio.1996.0292>
- 452 Bojilov, D., Dagnon, S., Ivanov, I. 2017. New insight into the flavonoid composition of
453 *Chenopodium botrys*. *Phytochem. Lett.*, 20, 316-321.
454 <https://doi.org/10.1016/j.phytol.2017.01.015>
- 455 Brand-Williams, W., Cuvelier, M.E., Berset, C. 1995. Use of a free radical method to evaluate
456 antioxidant activity. *LWT - Food Sci. Technol.*, 28, 25-30. [https://doi.org/10.1016/S0023-
457 6438\(95\)80008-5](https://doi.org/10.1016/S0023-6438(95)80008-5)
- 458 Cooper, M. A., Washburn K. W. 1998. The relationships of body temperature to weight gain, feed
459 consumption, and feed utilization in broilers under heat stress. *Poultry Sci.*, 77, 237-242.
460 <https://doi.org/10.1093/ps/77.2.237>
- 461 Dagnon S., Bojilov D., Docheva M., Edreva A. 2018. The relationship between main polyphenol
462 components and free radical scavenging activity of selected medicinal plants. *Int. J. Pharm.*
463 *Sci. Drug Res.*, 10, 131-138. [https://doi.org/ 10.25004/IJPSDR.2018.100305](https://doi.org/10.25004/IJPSDR.2018.100305)

- 464 Dimitrova, M., Ivanov, G., Mihalev, K., Slavchev, A., Ivanova, I., Vlaseva, R. 2019.
465 Investigation of antimicrobial activity of polyphenol-enriched extracts against probiotic lactic
466 acid bacteria. *Food Sci. Appl. Biotech.*, 2, 67-73. <https://doi.org/10.30721/fsab2019.v2.i1.57>
- 467 Dina E., Xynos N., Iliev H., Skaltsounis A.L., Aligiannis N. 2018. Exploitation of by-products
468 derived from hydrodistillation of rose (*Rosa damascena* Mill) petals. Proc. of 11th
469 International Symposium on Chromatography of Natural Products (ISCNP 2018), Lublin,
470 Poland.
- 471 Erbas, S., Baydar, H. 2016. Variation in scent compounds of oil-bearing rose (*Rosa damascena*
472 Mill.) produced by headspace solid phase microextraction, hydrodistillation and solvent
473 extraction. *Rec. Nat. Prod.*, 10, 555-565. [http://www.acgpubs.org/doc/2018080807064667-](http://www.acgpubs.org/doc/2018080807064667-RNP-1412-058.pdf)
474 [RNP-1412-058.pdf](http://www.acgpubs.org/doc/2018080807064667-RNP-1412-058.pdf)
- 475 Frankič, T., Voljč, M., Salobir, J., Rezar, V. 2009. Use of herbs and spices and their extracts in
476 animal nutrition. *Acta Argic. Slov.*, 94, 95-102. [http://aas.bf.uni-lj.si/zootehnika/94-](http://aas.bf.uni-lj.si/zootehnika/94-2009/PDF/94-2009-2-95-102.pdf)
477 [2009/PDF/94-2009-2-95-102.pdf](http://aas.bf.uni-lj.si/zootehnika/94-2009/PDF/94-2009-2-95-102.pdf)
- 478 Giusti, M.M., Jing, P. 2007. Analysis of anthocyanins. In: Carmen Socaciu Ed. *Food Colorants:*
479 *Chemical and Functional Properties*. 1st ed. CRC Press Taylor & Francis Group, Boca Raton,
480 FL, USA
- 481 Hashemi, S. R., Davoodi, H. 2001. Herbal plants and their derivatives as growth and health
482 promoters in animal nutrition. *Vet. Res. Commun.*, 35, 169-180.
483 <https://doi.org/10.1007/s11259-010-9458-2>
- 484 Jamroz, D., Wertelecki, T., Houszka, M., Kame, C. 2006. Influence of diet type on the inclusion
485 of plant origin active substances on morphological and histochemical characteristics of the

- 486 stomach and jejunum walls in chicken. *J. Anim. Physiol. Anim. Nutr.*, 90, 255-268.
487 <https://doi.org/10.1111/j.1439-0396.2005.00603.x>
- 488 Koch, W. 2019. Dietary polyphenols - important non-nutrients in the prevention of chronic
489 noncommunicable diseases. A systematic review. *Nutrients*, 11, 1039-1045.
490 <https://doi.org/10.3390/nu11051039>
- 491 Leenstra F. R., Pit R. 1987. Fat deposition in a broiler sire strain. 2. Comparisons among lines
492 selected for less abdominal fat, lower feed conversion ratio, and higher body weight after
493 restricted and ad libitum feeding. *Poultry Sci.*, 66, 193-202.
494 <https://doi.org/10.3382/ps.0660193>
- 495 Leman, A., O'Rourke, N., Hatcher, L., Stepanski, E. D. 2013. Chapter 8. JMP for Basic
496 Univariate and Multivariate Statistics: Methods for Researchers and Social Scientists, 2nd ed.
497 SAS Institute Inc., Cary, NC, USA.
- 498 Losinger, W. C. 1998. Feed-conversion ratio of finisher pigs in the USA. *Prev. Vet. Med.*, 36, 287-305.
499 [https://doi.org/10.1016/S0167-5877\(98\)00094-4](https://doi.org/10.1016/S0167-5877(98)00094-4)
- 500 Mollov, P., Mihalev, K., Shikov, V., Yoncheva, N., Karagyozov, V. 2007. Colour stability
501 improvement of strawberry beverage by fortification with polyphenolic copigments naturally
502 occurring in rose petals. *Innov. Food Sci. Emer. Technol.*, 8, 318-321.
503 <https://doi.org/10.1016/j.ifset.2007.03.004>
- 504 Oswell, N.J., Thippareddi, H., Pegg, R.B. 2018. Practical use of natural antioxidants in meat
505 products in the US: A review. *Meat Sci.*, 145, 469-479.
506 <https://doi.org/10.1016/j.meatsci.2018.07.020>
- 507 Özkan, G., Sagdiç, O., Baydar, N. G., Baydar, H. 2004. Note: Antioxidant and antibacterial
508 activities of *Rosa damascena* flower extracts. *Food Sci. Technol. Int.*, 10, 277-281.
509 <https://doi.org/10.1177/1082013204045882>

- 510 Pal, A., Bhushan, B., Narwal, R.K., Saharan V. 2018. Extraction and evaluation of antioxidant
511 and free radical scavenging potential correlated with biochemical components of red rose
512 petals. *Ir. J. Sci. Technol., Transac. A: Sci.*, 42, 1027–1036. [https://doi.org/10.1007/s40995-](https://doi.org/10.1007/s40995-016-0071-2)
513 [016-0071-2](https://doi.org/10.1007/s40995-016-0071-2)
- 514 Pasternak, H., Shalev B.A. 1983. Genetic-economic evaluation of traits in a broiler enterprise:
515 reduction of food intake due to increased growth rate. *Br. Poult. Sci.*, 24, 531-536.
516 <https://doi.org/10.1080/00071668308416772>
- 517 Pitman, R.S., Blumberg, R.S. 2000. First line of defense: The role of the intestinal epithelium as
518 an active component of the mucosal immune system. *J. Gastroent.*, 35, 805-814.
519 <https://doi.org/10.1007/s005350070017>
- 520 Rusanov, K., Garo, E., Rusanova, M., Fertig, O., Hamburger, M., Atanassov, I., Butterweck, V.
521 2014. Recovery of polyphenols from rose oil distillation wastewater using adsorption resins—a
522 pilot study. *Planta Med.*, 80, 1657-1664. <https://doi.org/10.1055/s-0034-1383145>
- 523 Schieber, A., Mihalev, K., Berardini, N., Mollov, P., Carlea, R. 2005. Flavonol glycosides from
524 distilled petals of *Rosa damascena* Mill. *Zeitschr. Naturforsch. C – J. Biosci.*, 60, 379-384.
525 <https://doi.org/10.1515/znc-2005-5-602>
- 526 Shikov, V., Kammerer, D., Mollov, P., Mihalev, K., Yoncheva, N., Carle, R. 2008. LC-MS
527 analysis and in vitro antioxidant activity of polyphenols from rose (*Rosa damascena* Mill.)
528 petals. *Plov. Uni. "P. Hilendarski" – Sci. Works - Chem. Proc. A*, 36, 59-63.[In Bulgarian]
529 https://blogs.uniplovdiv.net/argon/files/2008/03/007_NT36_2008.pdf
- 530 Shikov, V., Kammerer, D.R., Mihalev, K., Mollov, P., Carle, R. 2012. Antioxidant capacity and
531 colour stability of texture-improved canned strawberries as affected by the addition of rose

- 532 (Rosa damascena Mill.) petal extracts. Food Res. Int., 46, 552-556.
533 <https://doi.org/10.1016/j.foodres.2011.04.004>
- 534 Singleton, V.L., Rossi, J.A. 1965. Colorimetry of total phenolics with phosphomolybdic-
535 phosphotungstic acid reagents. Am. J. Enol. Viticul., 16, 144-158.
536 <http://www.ajevonline.org/content/16/3/144.full.pdf+html>
- 537 Slavov, A., Denev, P., Panchev, I., Shikov, V., Nenov, N., Yantcheva, N., Vasileva, I. 2017.
538 Combined recovery of polysaccharides and polyphenols from Rosa damascena wastes. Ind.
539 Crops Prod., 100, 85-94. <https://doi.org/10.1016/j.indcrop.2017.02.017>
- 540 Slavov, A., Panchev, I., Kovacheva, D., Vasileva, I. 2016. Physico-chemical characterization of
541 water-soluble pectic extracts from Rosa damascena, Calendula officinalis and Matricaria
542 chamomilla wastes. Food Hydrocol., 61, 469-476.
543 <https://doi.org/10.1016/j.foodhyd.2016.06.006>
- 544 Slavov, A., Vasileva, I., Stefanov, L., Stoyanova, A. 2017. Valorization of wastes from the rose
545 oil industry. Rev. Environ. Sci. Bio/Technol. 16, 309-325. [https://doi.org/10.1007/s11157-](https://doi.org/10.1007/s11157-017-9430-5)
546 [017-9430-5](https://doi.org/10.1007/s11157-017-9430-5)
- 547 Surai, P. F. 2014. Polyphenol compounds in the chicken/animal diet: from the past to the future.
548 J. Physiol. Anim. Nutr., 98,19-31. <https://doi.org/10.1111/jpn.12070>
- 549 Valenzuela-Grijalva, N. V., Pinelli-Saavedra, A., Muhlia-Almazan, A., Domínguez-Díaz, D.,
550 González-Ríos, H. 2017. Dietary inclusion effects of phytochemicals as growth promoters in
551 animal production. J. Ani. Sci. Technol., 59, 1-17. <https://doi.org/10.1186/s40781-017-0133-9>
- 552 Vasileva, I., Krastev, L., Slavov, A., Petkova, N., Yantcheva, N., Nenov, N., Krachmarov, A.,
553 Atanasova, A. 2019. Valorization of cocoa and rose waste for preparation of liqueurs. Food
554 Sci. Appl. Biotechnol., 2, 8-17. <https://doi.org/10.1016/10.30721/fsab2019.v2.i1.41>

- 555 Vlahova-Vangelova, D. B., Balev, D. K., Dragoev, S. G. 2018. Composition of foodstuff for
556 livestock and poultry. Certificate for registration of Utility model Reg. No 6 U1/October 15,
557 2018. Patent Office of the Republic of Bulgaria.
- 558 Windisch, W. M., Schedle, K., Plitzner, C., Kroismayr, A. 2008. Use of phytogenic products as
559 feed additives for swine and poultry. *J. Ani. Sci.*, 86, E140-E148.
560 <https://doi.org/10.2527/jas.2007-0459>
- 561 Yang, C., Chowdhury, M.A.K., Hou, Y., Gong, J. 2015. Phytogenic compounds as alternatives to in-feed
562 antibiotics: Potentials and challenges in application. *Pathogens*, 4, 137-156.
563 <https://doi.org/10.3390/pathogens4010137>

564 Table 1

565 Ingredients, chemical composition and energy values of basal diet

566

Components of combined feed for pigs	Grower for pigs, g/kg	Finisher for pigs, g/kg	Components of combined feed for lambs	Grower for lambs, g/kg	Finisher for lambs, g/kg	Components of combined feed for broilers	Starter for chicks, g/kg	Finisher for broilers, g/kg
Formulations								
Maize	150.00	130.00	Maize	450.00	250.00	Maize	400.00	450.00
Barley	250.00	100.00	Barley	-	150.00	Wheat	200.00	200.00
Wheat	270.00	500.00	Wheat	200.00	170.00	Soybean meal	150.00	150.00
Wheat bran	80.00	70.00	Oats	135.00	170.00	Sunflower pomace	100.00	100.00
Bio-concentrate BC14	250.00	-	Sunflower pomace	100.00	200.00	Vegetable fat	100.00	50.00
Bio-concentrate BC16	-	200.00	Vitamin-mineral premix	50.00	50.00	Vitamin-mineral premix including phosphate, lysine, methionine, enzymes, coccidiostatic, antitoxic substances	40.00	40.00
			Chalk	10.00	10.00	Chalk	7.00	7.00
			Alfalfa	100.00	100.00	Salt	3.00	3.00
Calculated chemical compositions, g/kg								
Crude protein	157.50	150.20	Crude protein	180.00	165.00	Crude protein	180.00	160.00
Crude fats	28.10	24.20	Crude fats	26.50	33.00	Crude fats	25.00	60.00
Crude ash	46.80	43.90	Crude ash	74.10	79.00	Crude ash	48.00	54.50
Dietary fibers	47.90	38.40	Dietary fibers	89.00	93.00	Dietary fibers	40.00	36.00
Amino acids, g/100g								
Lysine	0.80	0.72	Lysine	0.85	0.75	Lysine	0.92	0.88
Methionine			Methionine	0.40	0.33	Methionine	0.33	0.35
Minerals, g/100g								
Calcium	1.31	1.26	Calcium	1.30	1.50	Calcium	0.94	1.33
Phosphorus	0.85	0.31	Phosphorus	0.66	0.70	Phosphorus	0.78	0.82
			Sodium	0.40	0.54	Sodium	0.15	0.12
			Copper	0.0105	0.0095	Copper	0.0007	0.0010
			Magnesium	0.00011	-			
Energy intake, MJ/kg								
Digestible energy	13.46	13.72	Digestible energy	12.90	13.33	Digestible energy	12.87	13.80
Exchangeable energy	12.92	13.18	Exchangeable energy	12.34	12.82	Exchangeable energy	12.35	13.26

567

568 ^a The quantities of Siberian larch dihydroquercetin or dry distilled rose petals added as supplements to the
569 diets were calculated as 3.5 mg dihydroquercetin/kg/d (D1); 7.5 mg dihydroquercetin/kg/d (D2); 0.255 g dry
570 distilled rose petals/kg/d (R1); or 0.545 g dry distilled rose petals/kg/d (R2).

571 ^b The bio-concentrate BC14 contents: 312.10 g/kg crude protein, 10.70 g/kg crude fat, 153.00 g/kg crude ash,
572 38.10 g/kg crude fibers, 5.88 g/100 g lysine, 2.79 g/100 g methionine, 7.80 g/100 g calcium, 2.69 g/100 g

573 phosphorus, 268 mg/kg copperas sulphate, 670 mg/kg dl- α -tocopherol, 93800 UI/kg vitamin A, 16080 UI/kg
574 vitamin D3, 1975.845 kcal/kg total energy

575 ^c The bio-concentrate BC16 contents: 348.00 g/kg crude protein, 17.40 g/kg crude fat, 165.00 g/kg g crude
576 ash, 108.30 g/kg crude fibers, 2.26 g/100 g lysine, 0.67 g/100 g methionine, i.e. 1.25 g/100 g methionine +
577 cystine, 1.31 g/100 g threonine, 3.66 g/100 g calcium, 0.95 g/100 g phosphorus, i.e. 0.67 g/100 g absorbable
578 phosphorus, 0.78 g/100 g sodium, 560.00 mg/kg iron, 545.00 mg/kg zinc, 195.00 mg/kg manganese, 100.00
579 mg/kg copper, 4.10 mg/kg iodine, 1.50 mg/kg selenium, 0.40 g/100g antioxidants, 320.00 mg/kg vitamin E,
580 32500 UI/kg vitamin A, 6000 UI/kg vitamin D3.

581 ^d Vitamin-mineral premix for lambs contents: E672 Vitamin A (Retinal acetate) - 8000 IU/kg, Vitamin E (α -
582 Tocopherylacetat) - 40.00mg/kg, E671 Vitamin D3 (Cholecalciferol) - 2000 IU/kg, E1 Iron (Iron sulfate) -
583 170.26 mg/kg, E2 Iodine (Potassium iodate) - 1.10 mg/kg, E3 Cobalt (Cobalt carbonate) - 0.30 mg/kg, E4
584 Copper (Copper oxide) - 5.65 mg/kg, E5 Manganese (Manganese oxide) - 95.61 mg/kg, E6 Zinc (Zinc
585 oxide) - 108.86 mg/kg, E8 Selenium (Sodium selenite) - 0.40 mg/kg.

586

587 Table 2
 588 Content of polyphenolic compounds in dry rose petals, dry pressed distilled rose petals and waste water after
 589 isolation of essential oil from *Rosa damascena*
 590

Compounds	Waste products after isolation of essential oil from oil-bearing Bulgarian rose (<i>Rosa damascena</i>)			
	HPLC, t_R , min	Dry rose petals, mg/g	Dry pressed distilled rose petals, mg/g	Waste water, $\mu\text{g/ml}$
Gallic acid glycosides	2 - 20	$2,89 \pm 0,07$	$1,43 \pm 0,04$	$27,64 \pm 0,83$
Quercetin-3- <i>O</i> -galactoside	24.9 ± 0.6	1.64 ± 0.33	0.96 ± 0.18	19.72 ± 1.01
Quercetin 3- <i>O</i> -glucoside	25.5 ± 0.6	1.35 ± 0.21	0.79 ± 0.10	21.36 ± 0.56
Sum of Quercetin glycosides	24-26	$2,92 \pm 0,06$	$1,98 \pm 0,05$	$49,96 \pm 1,22$
Sum of Kaempferol glycosides	27-36	$2,81 \pm 0,06$	$1,97 \pm 0,05$	$47,53 \pm 1,19$
Kaempferol	43.5 ± 1.2	traces	0.22 ± 0.03	-

591
 592 ^a Gallic acid glycosides are determined as gallic acid, Sum of Quercetin glycosides as quercetin,
 593 and Sum of Kaempferol glycosides as kaempferol
 594
 595
 596
 597
 598
 599
 600
 601
 602
 603
 604
 605
 606
 607
 608
 609
 610
 611
 612
 613
 614
 615
 616

617 Table 3
 618 Chromatographic and spectral data of polyphenolic compounds in the ethylacetate and
 619 chloroform fractions of dry rose petals of *Rosa damascena*
 620

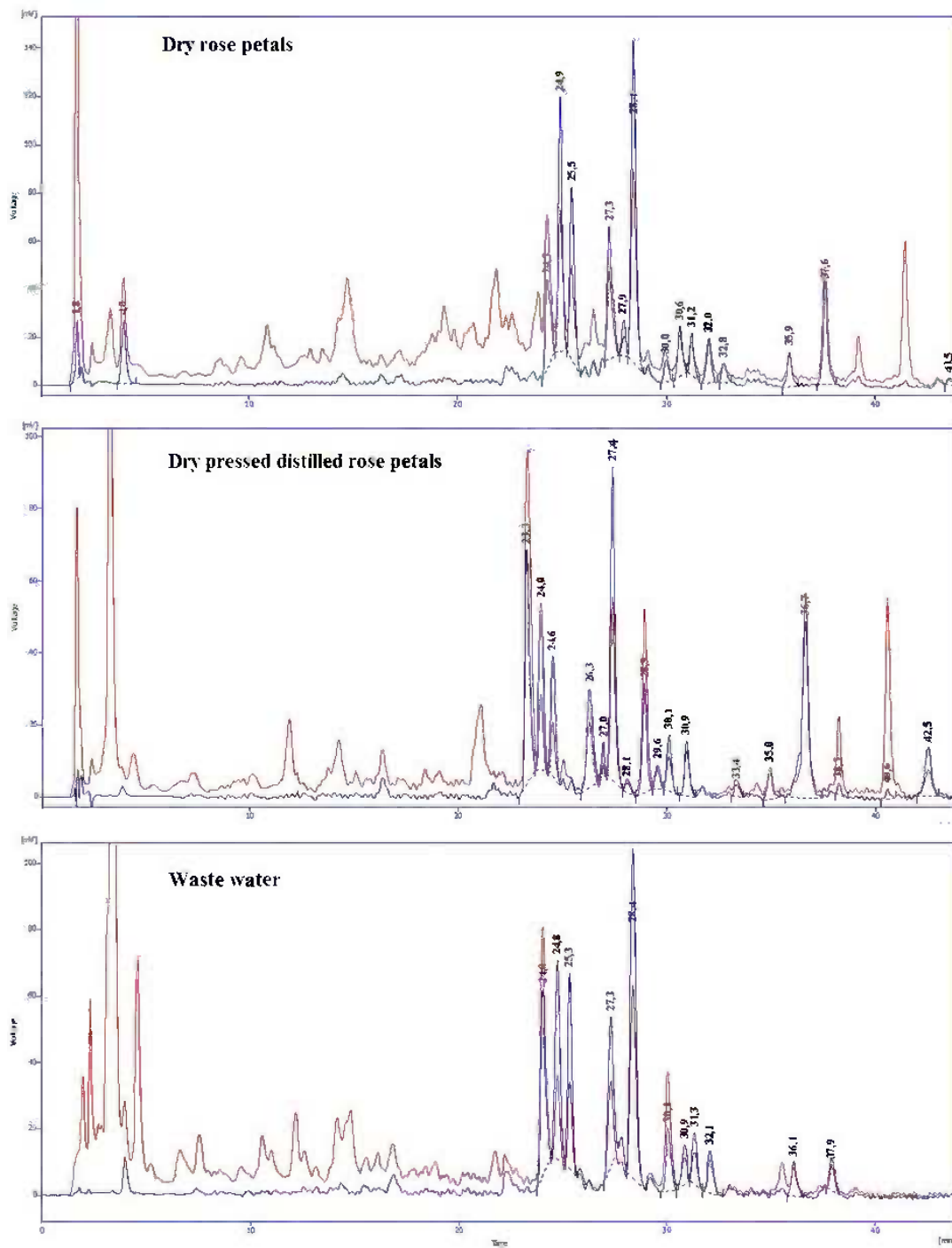
No	tr	Compounds	[M-H]-	MS/MS
1	1,79	Digalloyl hexoside	483	483, 331, 313, 169, 125, 107
2	2,71	Digalloyl hexoside	483	483, 331, 313, 169, 125, 107
3	3,48	Digalloyl hexoside	483	483, 331, 313, 169, 125, 107
4	5,17	Digalloyl hexoside	483	483, 331, 313, 169, 125, 107
5	5,93	Three galloyl hexoside	635	635, 483, 465, 313, 169, 125, 107
6	7,87	Methyl galloyl-galloyl hexoside	497	497, 447, 345, 334, 313, 183, 169
7	10,25	Quercetin galloyl hexoside	615	463, 301, 151
8	10,41	Quercetin galloyl hexoside	615	463, 301, 151
9	10,64	Quercetin 3- <i>O</i> -rutinoside	609	609, 301, 179
10	10,78	Quercetin 3- <i>O</i> -galactoside	463	463, 301, 300, 179, 151
11	10,99	Quercetin 3- <i>O</i> -glucoside	463	463, 301, 300, 179, 151
12	11,43	Quercetin galloyl hexoside	615	463, 301, 179, 151
13	11,49	Quercetin pentoside	433	433, 301
14a	11,74	Kaempferol hexoside	447	285, 151
14b		Kaempferol disaccharide	593	593, 447, 285, 151
15	11,92	Quercetin pentoside	433	433, 301 179, 151
16	12,15	Kaempferol hexoside	447	447, 285, 284, 151
17	12,18	Kaempferol hexoside	447	447, 285, 284, 151
18	12,24	Kaempferol hexoside	447	447, 285, 284, 151
19	12,64	Kaempferol galloyl hexoside	599	599, 447, 285, 151
20	12,93	Kaempferol pentoside	417	417, 285, 284, 151
21	13,08	Kaempferol disaccharide	593	593, 285
22	13,21	Kaempferol pentoside	417	417, 285, 179, 151
23	13,64	Kaempferol deoxyhexoside	431	431, 285
24	13,84	Quercetin acetyldisaccharide	651	651, 609, 301, 179, 151
25	14,66	Quercetin disaccharide	609	609, 463, 301, 179, 151
26	15,12	Kaempferol acetyldisaccharide	635	635, 593, 285, 257
27	15,93	Kaempferol disaccharide	593	593, 285
28	16,20	Kaempferol disaccharide	593	593, 285
29	15,38	Quercetin	301	301, 273, 179, 151, 121, 107
30	17,86	Kaempferol	285	285, 257, 213, 151, 107

622 Table 4
 623 Overall total feed consumption, average daily growth and feed-conversion ratio in experimental animals
 624

Parameters	Pigs			Lambs		Broilers	
	Control group (C)	Experimental group (R1)	Experimental group (R2)	Control group (C)	Experimental group (R)	Control group (C)	Experimental group (R)
Total feed consumption, kg	2.941±0.110 ^b	3.099±0.079 ^b	3.139±0.064 ^{bc}	2.182±0.044 ^a	2.172±0.057 ^a	6.780±0.101 ^d	6.980±0.114 ^d
Average daily growth, kg	0.717±0.062 ^c	0.828±0.051 ^c	0.911±0.036 ^{cd}	0.319±0.027 ^b	0.318±0.030 ^b	0.073±0.012 ^a	0.066±0.011 ^a
Feed-conversion ratio (FCR)	4.102 ± 0.291 ^c	3.743 ± 0.311 ^{bc}	3.446 ± 0.300 ^b	6.841± 0.672 ^d	6.831± 0.655 ^d	2.270± 0.334 ^a	2.220 ± 0.357 ^a

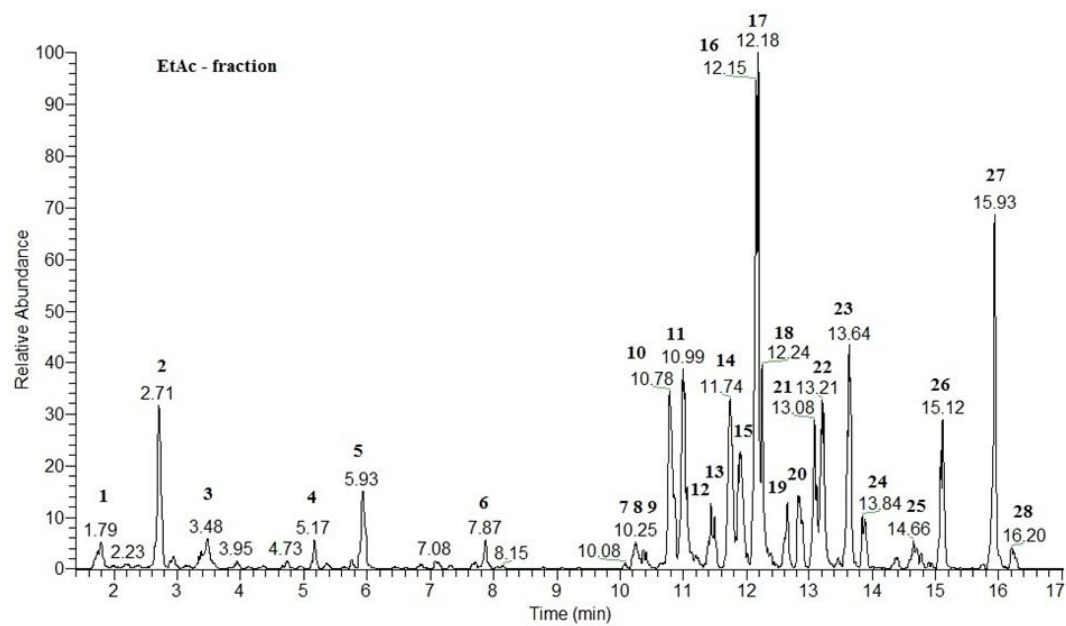
625
 626
 627

628 Fig 1. HPLC-PDA fingerprint profile of polyphenolic compounds of dry rose petals (*Rosa*
 629 *damascena*). Peak 4 – quercetin 3-*O*-galactoside ($t_R=24.9$ min), peak 5 – quercetin 3-*O*-
 630 glucoside ($t_R=25.5$ min), Peak 16- kaempferol ($t_R=43.5$); λ at 280nm and 352nm
 631
 632
 633
 634
 635



636

637 Fig. 2. Total ion current (TIC) of ethylacetate fraction of dry rose petals by UHPLC-MS/MS
638
639
640



641